

Introduction to R: Statistical Tools for Biologists

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◆ General introduction and information package

◆ Data manipulation and visualization

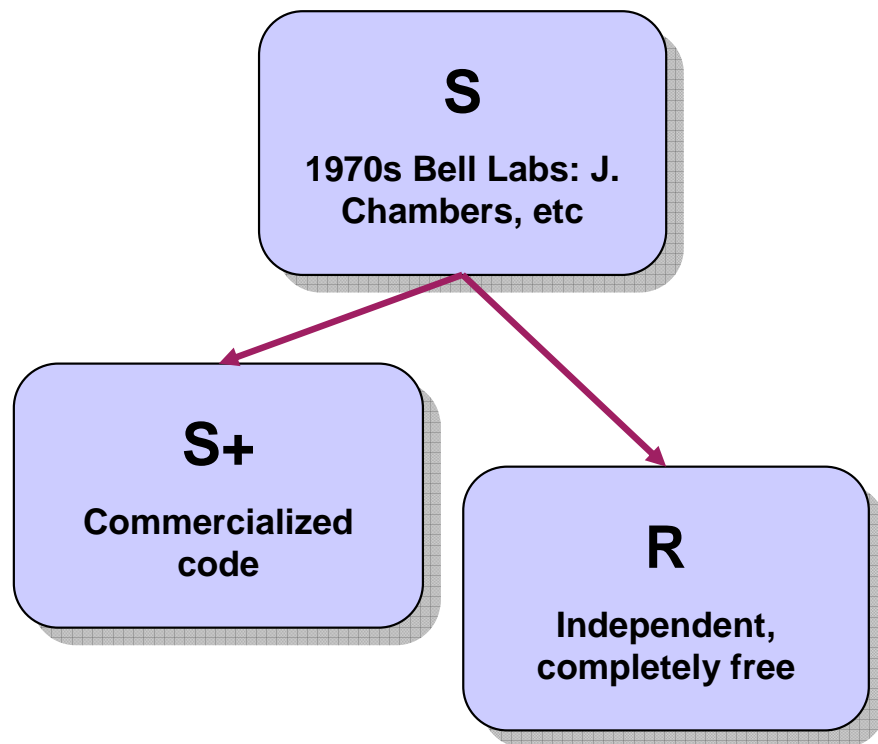
- ◆ basic operations
- ◆ import and export
- ◆ visualization

◆ Simple statistical analysis

- ◆ descriptive statistics
- ◆ statistical tests
- ◆ simple ANOVA (to be continued by Thomas)

◆ Demonstration of Bioconductor

History



R was created by
Ross Ihaka and **Robert Gentleman** at the
 University of Auckland, New Zealand, and is
 now developed by the
R Development Core Team.

Positive Features

- ◆ Scripting language of high level
- ◆ Interactive work with data (similar to MATLAB)
- ◆ Works under Windows, Linux, Mac OS
- ◆ Free code + verified algorithms
- ◆ Fast developing: new version each 2 month
- ◆ Extremely wide application: from biology to theoretical physics and computer sciences
- ◆ Very good information support
- ◆ Fast installation (with some exclusions)

Negative

- ◆ Not memory efficient (comparing to C)
- ◆ Slower then C/C++
- ◆ No advanced built-in GUI development tools

2. INFORMATION PACKAGE

Main Web-page:

cran.r-project.org

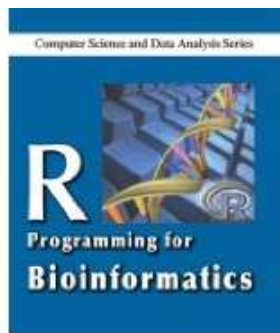
cran.r-project.org/manuals.html
cran.r-project.org/web/packages/
cran.r-project.org/other-docs.html

R-Project Seek Engine:

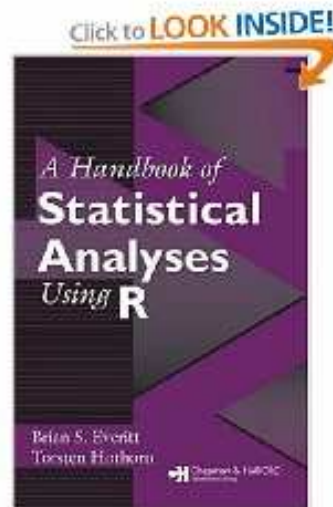
www.rseek.org

R/Bioconductor

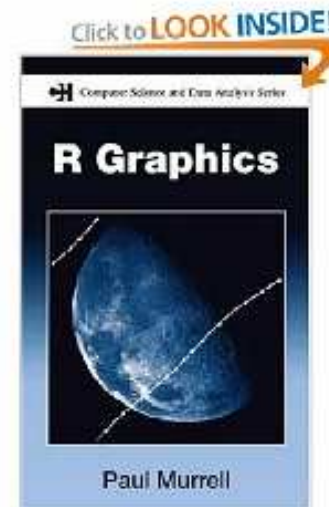
www.bioconductor.org



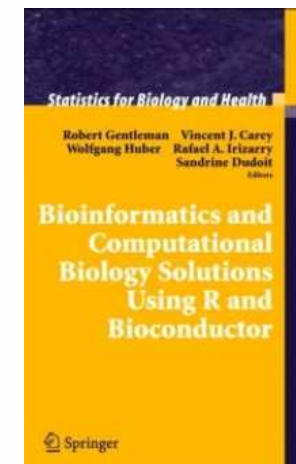
Robert Gentleman



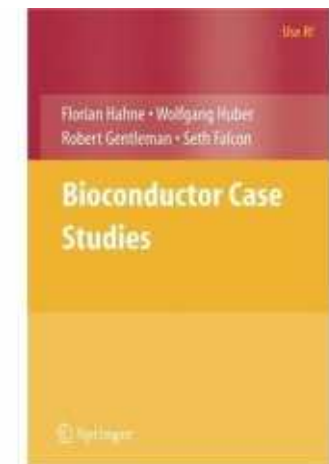
Peter S. Everitt
Torsten Hothorn



Paul Murrell



Springer

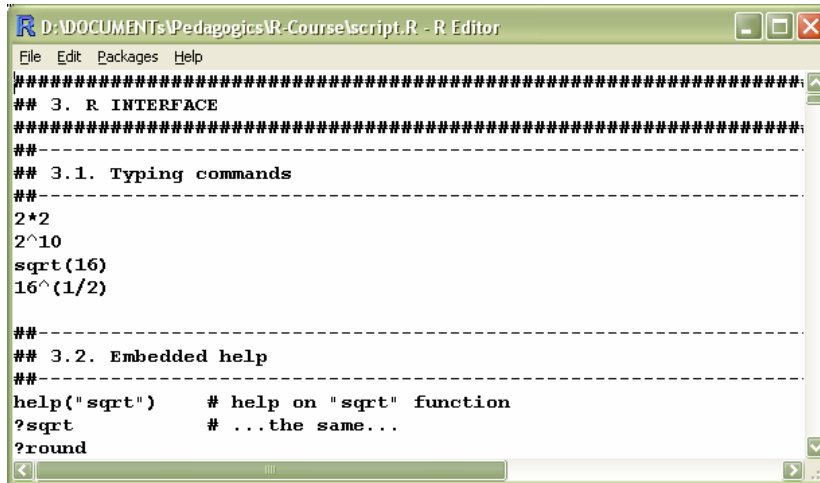


Springer

**All the materials and data used in the lecture
are available here**

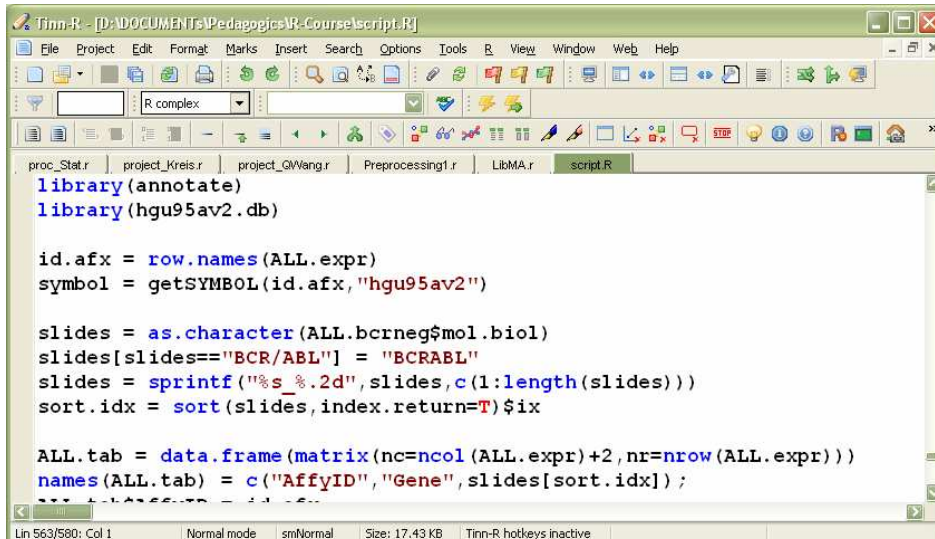
<http://edu.sablab.net/r>

Built-in Script Editor



```
D:\DOCUMENTS\Pedagogics\R-Course\script.R - R Editor
File Edit Packages Help
#####
## 3. R INTERFACE
#####
##-----
## 3.1. Typing commands
##-----
2*2
2^10
sqrt(16)
16^(1/2)
##-----
## 3.2. Embedded help
##-----
help("sqrt") # help on "sqrt" function
?sqrt       # ...the same...
?round
```

Alternative: Tinn-R
(highly recommended for win-users)



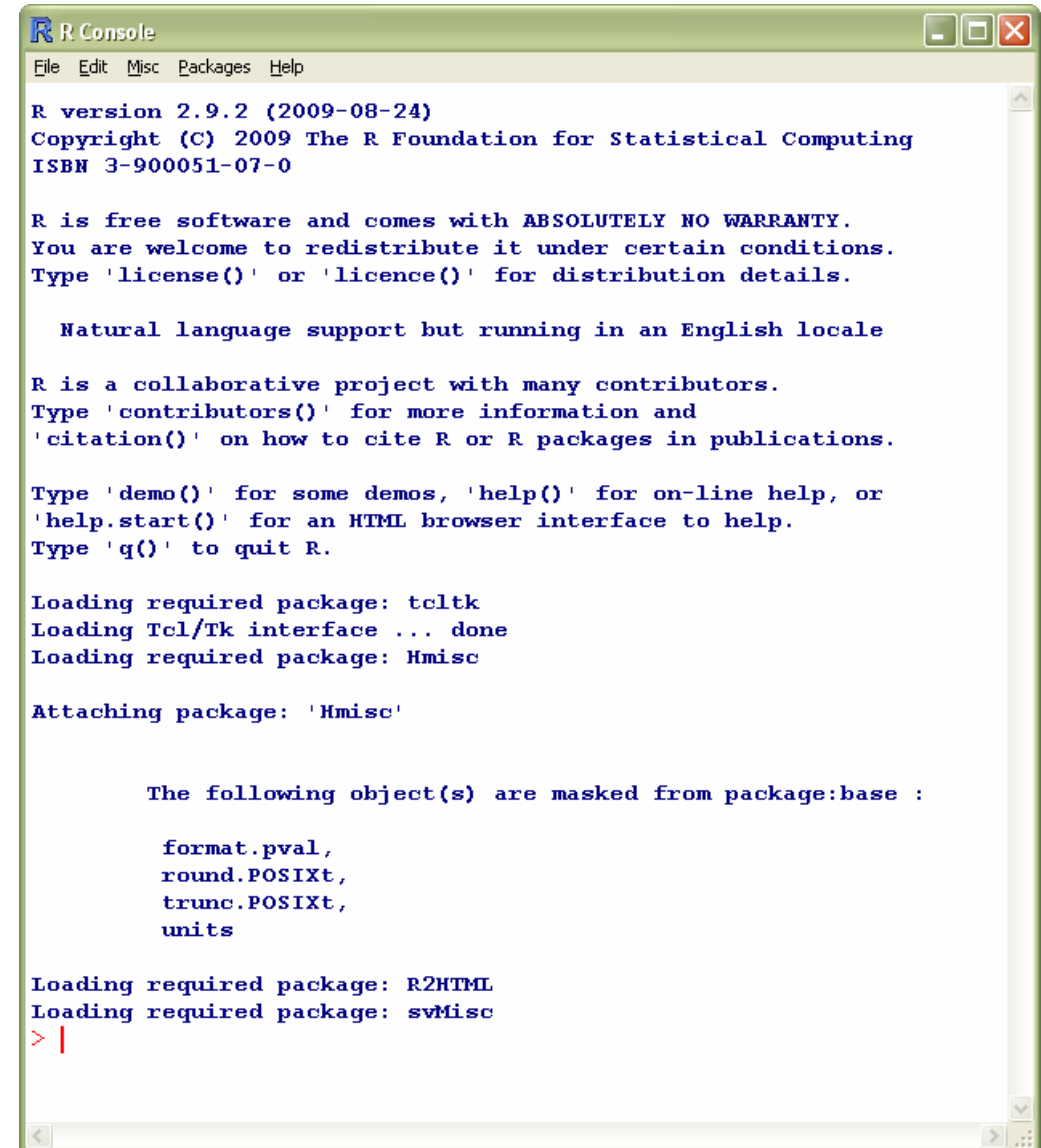
```
Tinn-R - [D:\DOCUMENTS\Pedagogics\R-Course\script.R]
File Project Edit Format Marks Insert Search Options Tools R View Window Web Help
R complex
proc_Stat.r | project_kreis.r | project_QWang.r | Preprocessing1.r | LibMA.r | script.R
library(annotate)
library(hgu95av2.db)

id.afx = row.names(ALL.expr)
symbol = getSYMBOL(id.afx, "hgu95av2")

slides = as.character(ALL.bcrneg$mol.biol)
slides[slides=="BCR/ABL"] = "BCRABL"
slides = sprintf("%s %.2d", slides, c(1:length(slides)))
sort.idx = sort(slides, index.return=T)$ix

ALL.tab = data.frame(matrix(nc=ncol(ALL.expr)+2, nr=nrow(ALL.expr)))
names(ALL.tab) = c("AffyID", "Gene", slides[sort.idx])
```

Console



```
R Console
File Edit Misc Packages Help

R version 2.9.2 (2009-08-24)
Copyright (C) 2009 The R Foundation for Statistical Computing
ISBN 3-900051-07-0

R is free software and comes with ABSOLUTELY NO WARRANTY.
You are welcome to redistribute it under certain conditions.
Type 'license()' or 'licence()' for distribution details.

Natural language support but running in an English locale

R is a collaborative project with many contributors.
Type 'contributors()' for more information and
'citation()' on how to cite R or R packages in publications.

Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.

Loading required package: tcltk
Loading Tcl/Tk interface ... done
Loading required package: Hmisc

Attaching package: 'Hmisc'

The following object(s) are masked from package:base :

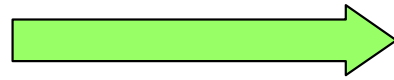
format.pval,
round.POSIXt,
trunc.POSIXt,
units

Loading required package: R2HTML
Loading required package: svMisc
> |
```

```
##-----
## 3.1. Typing commands
##-----
```

```
2*2
2^10
sqrt(16)
16^(1/2)
```

type in the
Console



```
> 2*2
[1] 4

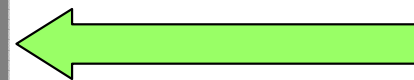
> 2^10
[1] 1024

> sqrt(16)
[1] 4

> 16^(1/2)
[1] 4

> |
```

select in Editor
and press Ctrl-R



```
2*2
2^10
sqrt(16)
16^(1/2)
```

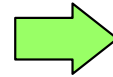
```
##-----
## 3.2. Embedded help
##-----
```

```
help("sqrt")      # help on "sqrt" function
?sqrt             # ...the same...
?round
??round          # fuzzy search for "round" in all help topics
apropos("plot")  # propose commands with the word "plot" inside the name

demo()           # show available demos
demo("image")    # start demo "image"
```

```
##-----
## 4.1. Mathematical operations and variables
##-----
```

```
x = 2
x
y <- 3
y
x + y -> z
#one more way to show the data
print(z)
#show variables in memory
ls()
```

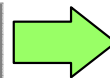


```
> x = 2
> x
[1] 2
> y <- 3
> y
[1] 3
> x + y -> z
> print(z)
[1] 5
> ls()
[1] "x" "y" "z"
```

See
"R Reference Card"
for more functions
and details

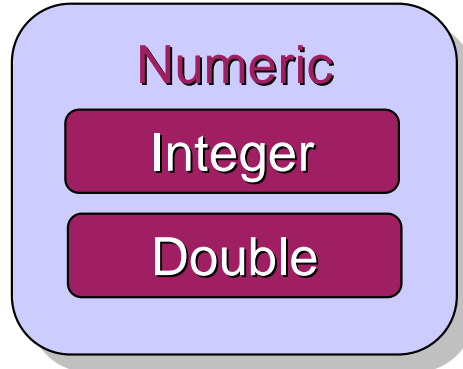
```
# remove all variables from memory
```

```
rm(list=ls())
ls()
```



```
> rm(list=ls())
> ls()
character(0)
>
```


Scalar Data



```
1
3.141593
```



```
TRUE
FALSE
```



```
"Hello, world!"
```



has a sense to use only in vectors or data frames

```
> answer=factor(c("yes", "no"))
> answer
[1] yes no
Levels: no yes
```

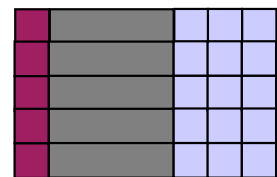
Data Containers



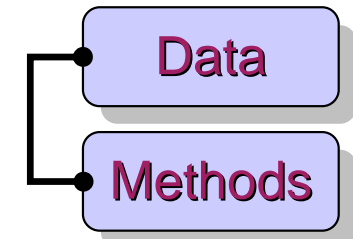
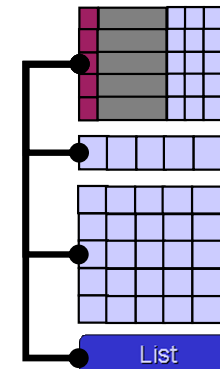
```
> x
[1] 1 2 3 4 5
```



```
> A
  [,1] [,2] [,3]
[1,]  1   3   5
[2,]  2   4   1
```



```
  name marks
1  Alex   10
2  Jean    8
3  David    7
```



4. BASIC OPERATIONS (4.2)

```
##-----
## 4.2. Types of data
##-----
```

```
##-----
## Numeric (integer, double)
i=5
i
i*2
i/2
i%/2 # integer division
i%%2 # remainder of integer division
round(1.5)
r=1.5
r
l=pi*2*r # let us calculate the circumference
l
```

```
##-----
## Boolean
b1=TRUE # try b1=T
b2=FALSE # try b2=F
b1 & b2 # logical AND
b1 | b2 # logical OR
!b1 # logical AND
xor(b1,b2) # logical XOR
r==1
r<1
```

For integer bitwise operation install and use
"bitops" package
bitAnd(), bitOr(), ...

```
> l=pi*2*r # let us calculate the
```

```
> l
[1] 9.424778
```

```
> b1=TRUE # try b1=T
```

```
> b2=FALSE # try b2=F
```

```
> b1 & b2 # logical AND
[1] FALSE
```

```
> b1 | b2 # logical OR
[1] TRUE
```

```
> !b1 # logical AND
[1] FALSE
```

```
> xor(b1,b2) # logical XOR
[1] TRUE
```

```
> r==1
[1] FALSE
```

```
> r<1
[1] TRUE
```

```
> |
```

```
##-----
## Character (strings)
st = "Hello, world!"
st
paste("We say:",st) # concatenation
sprintf("We say for the %d-rd time: %s..",3,st) # a more powerfull method a-la C
sprintf("By the way pi=%f, and e=%f",pi,exp(1))

sub(", world","",st) # replace a part of the sting
# (*) in R the regular expression are used to define the patte
casefold(st, upper=T) # change the case
nchar(st) # number of characters
strsplit(st,"")[[1]] # (*) transforms a string into the vector of single characters
```

```
> st
[1] "Hello, world!"

> paste("We say:",st) # concatenation
[1] "We say: Hello, world!"

> sprintf("We say for the %d-rd time: %s..",3,st) # a more powerfu$
[1] "We say for the 3-rd time: Hello, world!.."

> sprintf("By the way pi=%f, and e=%f",pi,exp(1))
[1] "By the way pi=3.141593, and e=2.718282"

> sub(", world","",st) # replace a part of the sting
[1] "Hello!"
```

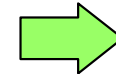
```
## how to check who is who?
is.character(st)
is.numeric(st)
is.numeric(1)
```

```

##-----
## 4.3. Special values
##-----
## NA - Not-Available (missing data)
na = NA
na + 1
100>na
na==na
is.na (na)

## Inf - Infinity (+/- infinite data)
0*1/0
-1/0
is.infinite(1/0)

## NaN - Not-A-Number
0/0
is.nan(sqrt(-1))
  
```



```

> na = NA
> na + 1
[1] NA
> 100>na
[1] NA
> na==na
[1] NA
> is.na(na)
[1] TRUE
> 0*1/0
[1] NaN
> -1/0
[1] -Inf
> is.infinite(1/0)
[1] TRUE
> 0/0
[1] NaN
> is.nan(sqrt(-1))
[1] TRUE
Warning message:
In sqrt(-1) : созданы NaN
> |
  
```

```
##-----
## 4.4. Vectors, matrixes and lists
##-----
```

```
## Vector creation
a = c(1,2,3,4,5)
a
a[1]+a[4]

b=5:9
a+b  ##(*) try b=5:10. Can you explain the effect?

seq(from=1,to=10,by=0.5) #sequence
rep(1:4, 2)             # same as rep(1:4, times=2)
rep(1:4, each=2)       # not the same

txt = c(st, "Let's try vectors", "bla-bla-bla")
txt

boo = c(T,F,T,F,T)
boo

## Vector indexes
a
a[1:3] # take a part of vector by index numbers
a[boo] # take a part of vector by logical vector
a[a>2] # take a part by a condition (!!!)
```

```
> a = c(1,2,3,4,5)
> a
[1] 1 2 3 4 5
> a[1]+a[4]
[1] 5
> b=5:9
> a+b  ##(*) try b=5:10. Can you explain the effect
[1] 6 8 10 12 14
> seq(from=1,to=10,by=0.5) #sequence
[1] 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5 5.0
[11] 6.0 6.5 7.0 7.5 8.0 8.5 9.0 9.5 10.0
> rep(1:4, 2)             # same as rep(1:4, times=2)
[1] 1 2 3 4 1 2 3 4
> rep(1:4, each=2)       # not the same
[1] 1 1 2 2 3 3 4 4
> txt = c(st, "Let's try vectors", "bla-bla-bla")
> txt
[1] "Hello, world!"      "Let's try vectors"
[3] "bla-bla-bla"
> boo = c(T,F,T,F,T)
> boo
[1] TRUE FALSE TRUE FALSE TRUE
> a
[1] 1 2 3 4 5
> a[1:3] # take a part of vector by index numbers
[1] 1 2 3
```

```

##-----
## Matrix
A=matrix(0,nrow=5, ncol=5)
A
A=A-1 # add scalar
A
A=A+a # add vector
A
t(A) #
B=A+t(A) #
B
B*B #
B%*%B #
  
```

```

> Data
      name    sex weight age survival code
1 Mouse_1  Male   21  160     TRUE    0
2 Mouse_2 Female   17  131    FALSE    1
3 Mouse_3 Female   20  149     TRUE    2
4 Mouse_4  Male   22  187    FALSE    3
5 Mouse_5  Male   19  141     TRUE    4
> |
  
```

```

##-----
## Data frame
Data=data.frame(A) # alternatively: D=data.frame(matrix(nr=5,nc=5))
Data
## let us add a column to D
mice = sprintf("Mouse_%d",1:5)
Data=cbind(mice,Data)
## put the names to the variables
names(Data)=c("name","sex","weight","age","survival","code")
Data
## put in the data manually
Data$name=sprintf("Mouse_%d",1:5)
Data$sex=c("Male","Female","Female","Male","Male")
Data$weight=c(21,17,20,22,19)
Data$age=c(160,131,149,187,141)
Data$survival=c(T,F,T,F,T)
Data
  
```

4. BASIC OPERATIONS (4.4)

```

## (!!!) see the structure of the objects
str(Data)

## see the structure of the objects
head(Data)

## summary on the data
summary(Data)

## Let's use factors
Data$sex = factor(Data$sex)
summary(Data)

## usefull commands when working with factors:
levels(Data$sex)      # returns levels of the factor
nlevels(Data$sex)    # returns number of levels
as.character(Data$sex) # transform into strings
  
```

```

> Data
  name      sex weight age survival code
1 Mouse_1  Male    21  160     TRUE    0
2 Mouse_2 Female    17  131     FALSE    1
3 Mouse_3 Female    20  149     TRUE     2
4 Mouse_4  Male    22  187     FALSE    3
5 Mouse_5  Male    19  141     TRUE     4
> |
  
```

```

> str(Data)
'data.frame':   5 obs. of  6 variables:
 $ name      : chr  "Mouse_1" "Mouse_2" "
 $ sex       : chr  "Male" "Female" "Fema
 $ weight    : num  21 17 20 22 19
 $ age       : num  160 131 149 187 141
 $ survival  : logi  TRUE FALSE TRUE FALS
 $ code      : num  0 1 2 3 4
  
```

```

> head(Data)
  name      sex weight age survival co
1 Mouse_1  Male    21  160     TRUE
2 Mouse_2 Female    17  131     FALSE
3 Mouse_3 Female    20  149     TRUE
4 Mouse_4  Male    22  187     FALSE
5 Mouse_5  Male    19  141     TRUE
  
```

```

> summary(Data)
  name      sex
Length:5      Length:5
Class :character  Class :character
Mode :character  Mode :character
  
```

```

  age      survival      coc
Min.   :131.0  Mode :logical  Min.   :
1st Qu.:141.0  FALSE:2      1st Qu.:
Median :149.0  TRUE :3       Median :
Mean   :153.6  NA's :0       Mean   :
3rd Qu.:160.0  3rd Qu.:
Max.   :187.0  Max.   :
  
```

```

##-----
## Lists

L=list()
L$Data=Data
L$descr = "A fake experiment with virtual mice"
L$num = nrow(Data)
str(L)

## how to access the fields? Simple!
L$Data # try also L$"Data"
L$num
## or
L[[1]]
L[[3]]
  
```

```

## clear all
ls()
rm(list=ls())
ls()
  
```

```

> str(L)
List of 3
 $ Data : 'data.frame': 5 obs. of 6 variables
  ..$ name      : chr [1:5] "Mouse_1" "Mouse_2" "Mouse_3" "Mouse_4" "Mouse_5"
  ..$ sex       : Factor w/ 2 levels "Female", "Male": 2 1 1 2 2
  ..$ weight    : num [1:5] 21 17 20 22 19
  ..$ age       : num [1:5] 160 131 149 187 141
  ..$ survival  : logi [1:5] TRUE FALSE TRUE FALSE TRUE
  ..$ code      : num [1:5] 0 1 2 3 4
 $ descr: chr "A fake experiment with virtual mice"
 $ num  : int 5
  
```

```

> L$Data # try also L$"Data"
  name sex weight age survival code
1 Mouse_1 Male 21 160 TRUE 0
2 Mouse_2 Female 17 131 FALSE 1
3 Mouse_3 Female 20 149 TRUE 2
4 Mouse_4 Male 22 187 FALSE 3
5 Mouse_5 Male 19 141 TRUE 4
  
```

```

> L$num
[1] 5
  
```

```

> L[[1]]
  name sex weight age survival code
1 Mouse_1 Male 21 160 TRUE 0
2 Mouse_2 Female 17 131 FALSE 1
3 Mouse_3 Female 20 149 TRUE 2
4 Mouse_4 Male 22 187 FALSE 3
5 Mouse_5 Male 19 141 TRUE 4
  
```

```

> L[[3]]
[1] 5
  
```



```

##-----
## "Currency" dataset: learn how to use "header" and "as.is"
##-----
Currency = read.table("http://edu.sablab.net/r/data/currency.txt")
str(Currency)
head(Currency)
## hmm.. wrong... let's ask to add header
Currency = read.table("http://edu.sablab.net/r/data/currency.txt",
                      header=T)

str(Currency)
head(Currency)
## still, it's better to consider dates as strings, not as factors
Currency = read.table("http://edu.sablab.net/r/data/currency.txt",
                      header=T, as.is=T)

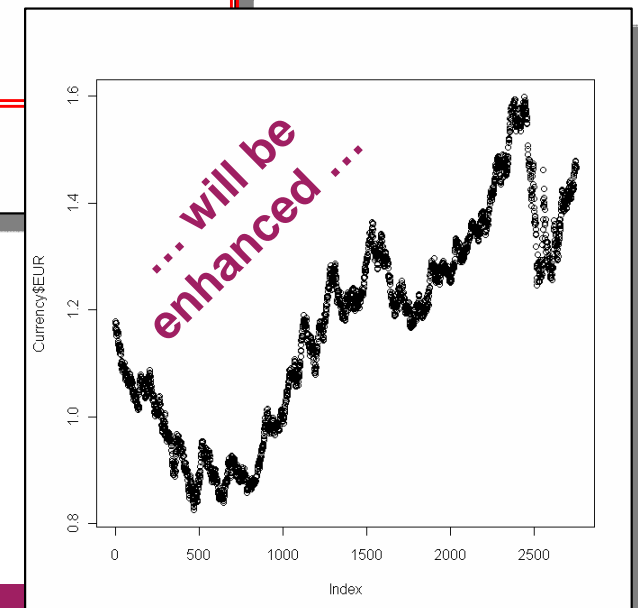
str(Currency)
summary(Currency)
## correct!
plot(Currency$EUR)

```

```

> str(Currency)
'data.frame': 2749 obs. of 2 variables:
 $ Date: chr "1999-01-04" "1999-01-05" "1999-01-06" "1999-01-07" ...
 $ EUR : num 1.18 1.18 1.17 1.16 1.17 ...

```



```

##-----
## 5.2. "Shop" dataset: learn how to use "sep"
##-----
Shop = read.table("http://edu.sablab.net/r/data/shop.txt",
                  header=T)
## hm... an error!
Shop = read.table("http://edu.sablab.net/r/data/shop.txt",
                  header=T, sep="\t")
## correct!
str(Shop)
head(Shop)

```

```

> summary(Shop)

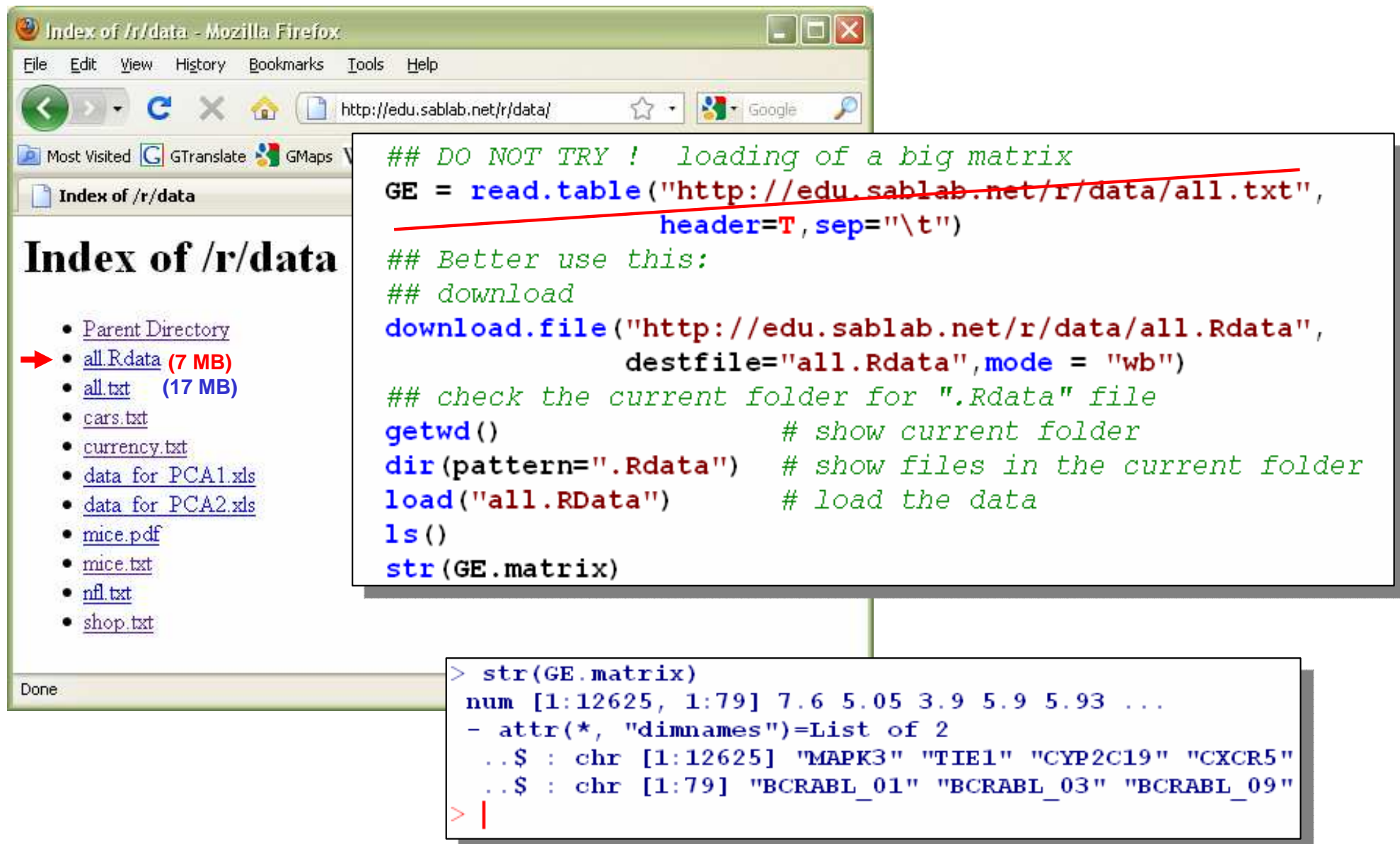
```

Customer		Payment		Items		Discount	
Min.	: 1.00	American Express:	2	Min.	: 1.00	Min.	: 0.00
1st Qu.:	25.75	Discover	: 4	1st Qu.:	1.00	1st Qu.:	0.00
Median	: 50.50	Mastercard	:14	Median	: 2.00	Median	: 15.00
Mean	: 50.50	Proprietary Card:	70	Mean	: 3.22	Mean	: 22.45
3rd Qu.:	75.25	Visa	:10	3rd Qu.:	4.00	3rd Qu.:	31.25
Max.	:100.00			Max.	:17.00	Max.	:158.30
Sales		Gender		Status		Age	
Min.	: 13.23	Female:	93	Married:	84	Min.	:20.00
1st Qu.:	39.60	Male	: 7	Single	:16	1st Qu.:	32.00
Median	: 59.70					Median	:42.00
Mean	: 77.60					Mean	:43.08
3rd Qu.:	100.90					3rd Qu.:	50.00
Max.	:287.59					Max.	:78.00

```

> |

```



The screenshot shows a Mozilla Firefox browser window with the address bar at `http://edu.sablab.net/r/data/`. The page title is "Index of /r/data" and the content shows a directory listing with links to various files. A red arrow points to the link `all.Rdata (7 MB)`. Overlaid on the browser is a terminal window containing R code for downloading and loading data. A red line is drawn through the `read.table` function in the code.

```

## DO NOT TRY ! loading of a big matrix
GE = read.table("http://edu.sablab.net/r/data/all.txt",
      header=T, sep="\t")
## Better use this:
## download
download.file("http://edu.sablab.net/r/data/all.Rdata",
             destfile="all.Rdata", mode = "wb")
## check the current folder for ".Rdata" file
getwd()           # show current folder
dir(pattern=".Rdata") # show files in the current folder
load("all.RData") # load the data
ls()
str(GE.matrix)

```

```

> str(GE.matrix)
num [1:12625, 1:79] 7.6 5.05 3.9 5.9 5.93 ...
- attr(*, "dimnames")=List of 2
 ..$ : chr [1:12625] "MAPK3" "TIE1" "CYP2C19" "CXCR5"
 ..$ : chr [1:79] "BCRABL_01" "BCRABL_03" "BCRABL_09"
> |

```

5. DATA IMPORT AND EXPORT (5.4)

```
##-----  
## 5.4. Data export  
##-----  
write.table(Shop, "shop.txt", sep = "\t",  
            eol = "\n", na = "NA", dec = ".",  
            row.names = F,  
            qmethod = c("escape", "double"));  
  
save(Shop, file="shop.Rdata")  
  
getwd()  
dir()  
## if you need to set working folder, use  
setwd("...put here desired path...")  
  
## clear all  
rm(list=ls())
```

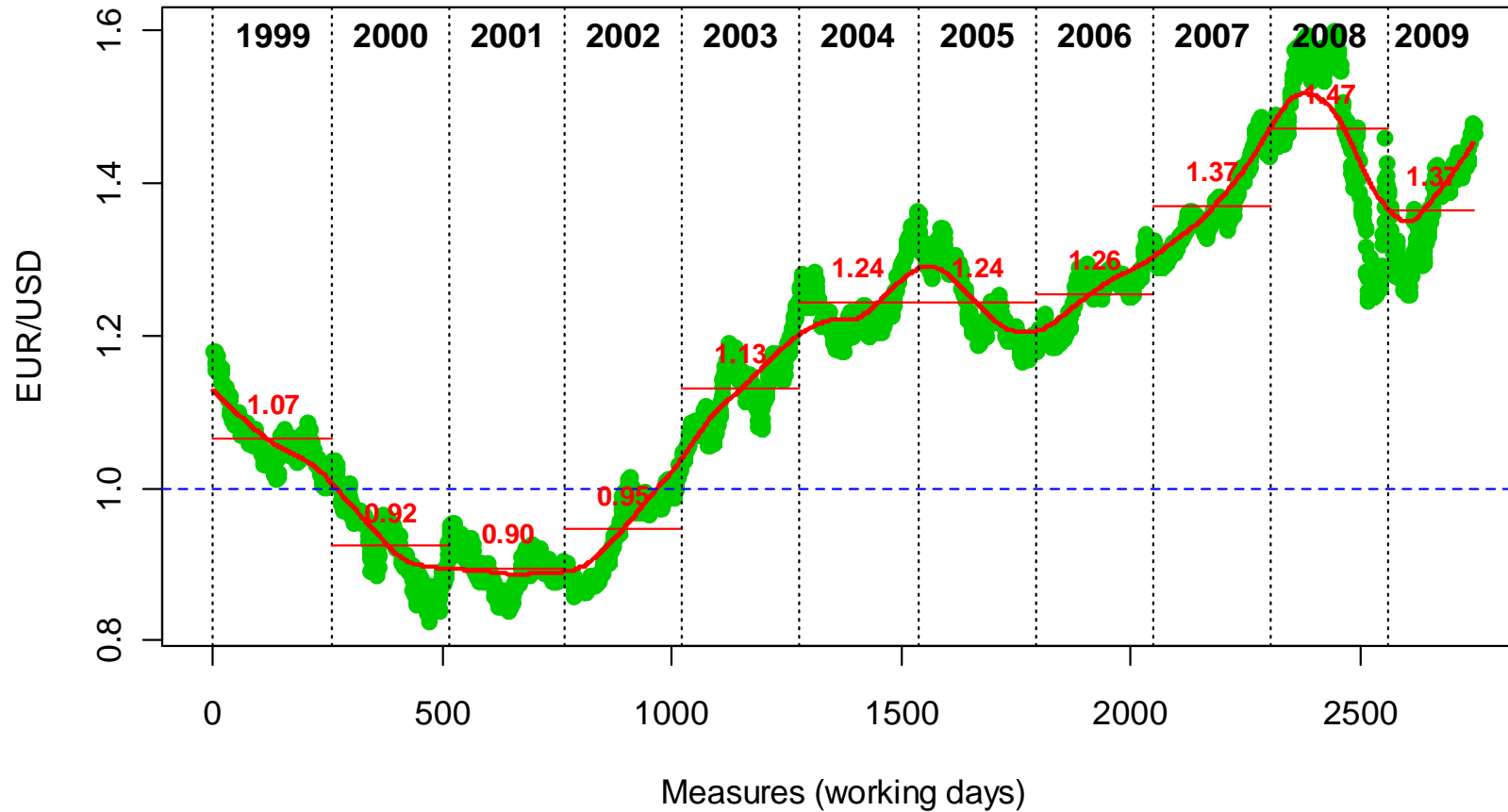
```
> write.table(Shop, "shop.txt", sep = "\t",  
+            eol = "\n", na = "NA", dec = ".",  
+            row.names = F,  
+            qmethod = c("esc ..." ... [TRUNC  
  
> save(Shop, file="shop.Rdata")  
  
> getwd()  
[1] "D:/DOCUMENTs/Pedagogics/R-Course"  
  
> dir()  
[1] "all.Rdata"  
[2] "Data"  
[3] "Handout"  
[4] "Info"  
[5] "Introduction to R.doc"  
[6] "Nazarov_091009_RCourse.ppt"  
[7] "plan.doc"  
[8] "script.R"  
[9] "shop.Rdata"  
[10] "shop.txt"  
> |
```

```
##-----
## 6.1. Plot time-series and smooth
##-----
## initiate window
windows(8,5) # try x11()
## get data
Currency = read.table("http://edu.sablab.net/r/data/currency.txt",
                      header=T, as.is=T)
## plot the currency behaviour for the last 10 years
plot(Currency$EUR)
## let's make it more beautiful
windows(8,5)
plot(Currency$EUR, col=3, pch=19,
     main="EUR/USD ratio for 11 years",
     ylab="EUR/USD",
     xlab="Measures (working days)")
## add smoothing. Try different "f"
smooth = lowess(Currency$EUR, f=0.1)
lines(smooth, col=2, lwd=2)
## add 1 level
abline(h=1, col=4, lty=2)
```

```
## (*) add years
year=1999 # an initial year
while (year<=2009){ # loop for all the years up to now
  idx=grep(paste("^", year, sep=""), Currency$Date) # take the indexes of the meas
  average=mean(Currency$EUR[idx]) # calculate the average ratio for the "year"
  abline(v=min(idx), col=1, lty=3) # draw the year separator
  lines(x=c(min(idx), max(idx)), y=c(average, average), col=2) # draw the average r
  text(median(idx), max(Currency$EUR), sprintf("%d", year), font=2) # write the year
  text(median(idx), average+0.05, sprintf("%.2f", average), col=2, font=2, cex=0.8) #
  year=year+1;
}
```

```
> windows(8,5) # try x11()
> Currency = read.table("http://edu.sablab.net/r/data/currency.txt",
+                       header=T, as.is=T)
> plot(Currency$EUR)
> windows(8,5)
> plot(Currency$EUR, col=3, pch=19,
+      main="EUR/USD ratio for 11 years",
+      ylab="EUR/USD",
+      xlab="Measures (working days)")
> smooth = lowess(Currency$EUR, f=0.1)
> lines(smooth, col=2, lwd=2)
> abline(h=1, col=4, lty=2)
> year=1999 # an initial year
> while (year<=2009){ # loop
+   idx=grep(paste("^", year, sep=""), Currency$Date)
+   average=mean(Currency$EUR[idx])
+   abline(v=min(idx), col=1, lty=3)
+   lines(x=c(min(idx), max(idx)), y=c(average, average), col=2)
+   text(median(idx), max(Currency$EUR), sprintf("%d", year), font=2)
+   text(median(idx), average+0.05, sprintf("%.2f", average), col=2, font=2, cex=0.8)
+   year=year+1;
> }
```

EUR/USD ratio for 11 years



```

##-----
## 6.2. 3D visualization and custom functions
##-----
## define the function to be plotted
sinc = function(x) {
  y = sin(x)/x
  y[is.na(y)]=1
  return(y)
}
rotsinc = function(x,y) {
  10*sinc( sqrt(x^2+y^2) )
}
## generate values
x = seq(-10, 10, length = 50)
y = x
z = outer(x, y, rotsinc)
windows()
## plot 3D image rotated by "theta" and "phi"
persp(x, y, z, theta = 30, phi = 30, expand = 0.5,
      col = "lightblue", ltheta = 120, shade = 0.75,
      ticktype = "detailed",
      xlab = "X", ylab = "Y", zlab = "Z")
## put the title and subscript
title(main = expression(z == sinc(sqrt(x^2 + y^2))),
      sub="Note: sinc(x) = sin(x) / x")
  
```

```

> sinc = function(x) {
+   y = sin(x)/x
+   y[is.na(y)]=1
+   return(y)
+ }

> rotsinc = function(x,y) {
+   10*sinc( sqrt(x^2+y^2) )
+ }

> x = seq(-10, 10, length = 50)

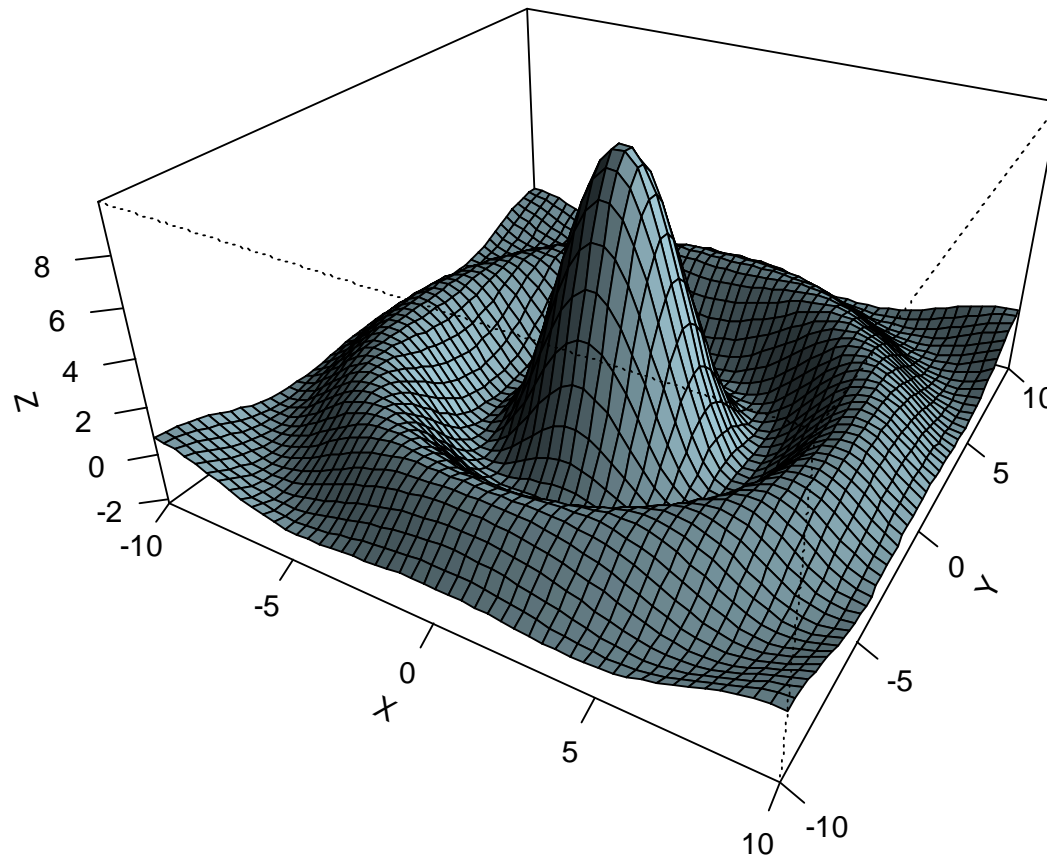
> y = x

> z = outer(x, y, rotsinc)

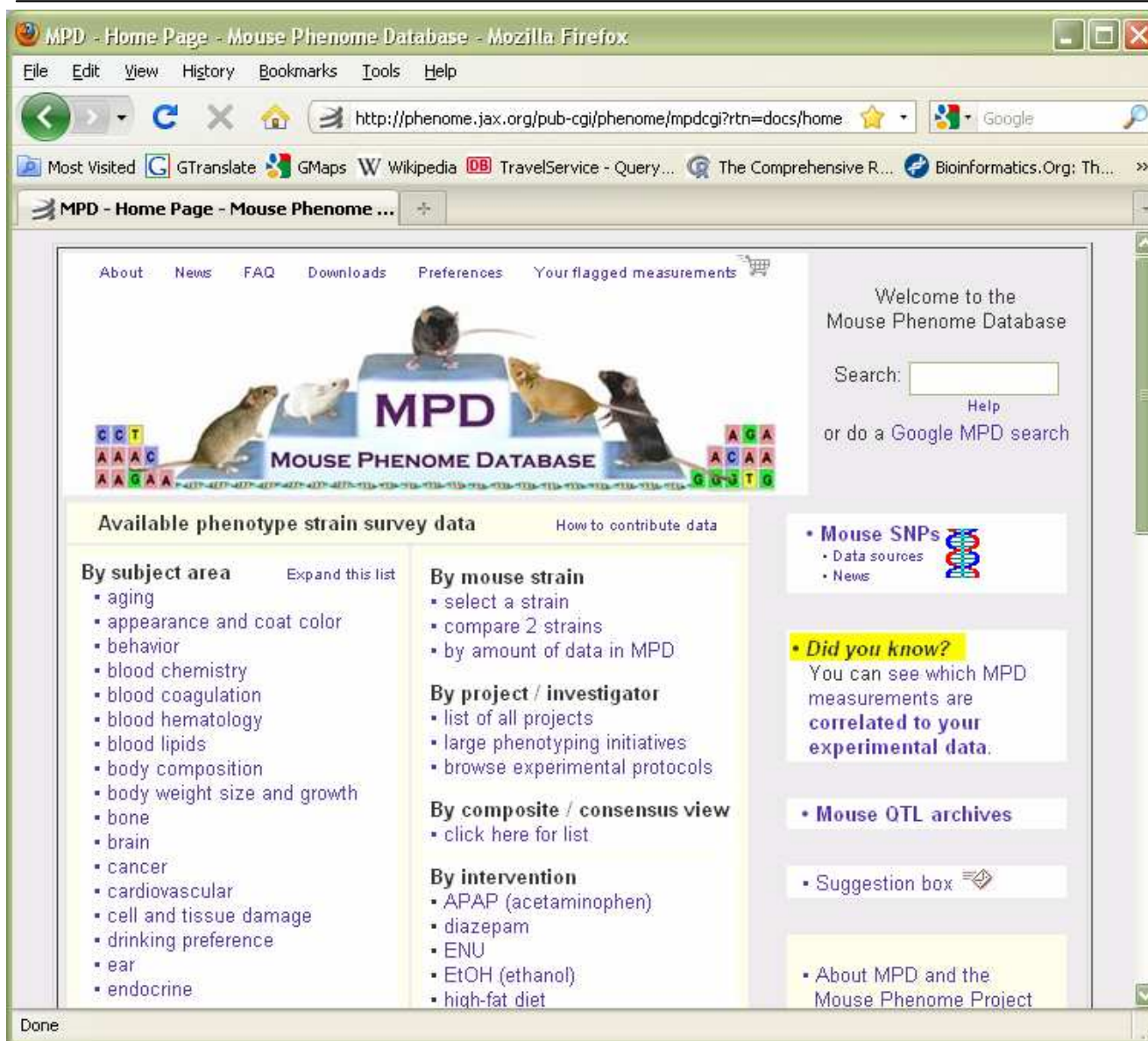
> persp(x, y, z, theta = 30, phi = 30, expand = 0.5,
+       col = "lightblue", ltheta = 120, shade = 0.75,
+       ticktype = "detailed",
+       xlab = "X", ylab = "Y", zlab = "Z")

> title(main = expression(z == sinc(sqrt(x^2 + y^2))),
+       sub="Note: sinc(x) = sin(x) / x")
> |
  
```

$$z = \text{sinc}(\sqrt{x^2 + y^2})$$



Note: $\text{sinc}(x) = \sin(x) / x$



MPD - Home Page - Mouse Phenome Database - Mozilla Firefox

File Edit View History Bookmarks Tools Help

http://phenome.jax.org/pub/cgi/phenome/mpdcgi?rtn=docs/home

MPD - Home Page - Mouse Phenome ...

About News FAQ Downloads Preferences Your flagged measurements

Welcome to the Mouse Phenome Database

Search:

Help

or do a Google MPD search

Available phenotype strain survey data [How to contribute data](#)

By subject area [Expand this list](#)

- aging
- appearance and coat color
- behavior
- blood chemistry
- blood coagulation
- blood hematology
- blood lipids
- body composition
- body weight size and growth
- bone
- brain
- cancer
- cardiovascular
- cell and tissue damage
- drinking preference
- ear
- endocrine

By mouse strain

- select a strain
- compare 2 strains
- by amount of data in MPD

By project / investigator

- list of all projects
- large phenotyping initiatives
- browse experimental protocols

By composite / consensus view

- click here for list

By intervention

- APAP (acetaminophen)
- diazepam
- ENU
- EtOH (ethanol)
- high-fat diet

Mouse SNPs

- Data sources
- News

Did you know?

You can see which MPD measurements are **correlated to your experimental data.**

Mouse QTL archives

Suggestion box

About MPD and the Mouse Phenome Project

Tordoff MG, Bachmanov AA

Survey of calcium & sodium intake and metabolism with bone and body composition data

Project symbol: **Tordoff3**

Accession number: **MPD:103**

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- [shop.txt](#)

```

##-----
## 6.3. Mouse phenom :)
##-----
## load data
Mice=read.table("http://edu.sablab.net/r/data/mice.txt",
                header=T,as.is=F)

str(Mice)
## initiate window
windows(10,8)
par(mfrow=c(2,2))
## plot a factorial data
plot(Mice$strain,las=2,
      col=rainbow(nlevels(Mice$strain)),cex.names =0.7)
title("Number of mice from each strain")
## plot factorial data as pie
pie(summary(Mice$sex), col=c("pink","lightblue"))
title("Gender composition (f:female, m:male)")
## try to use special command "barplot" as well
## a histogram
hist(Mice$bw_start,probability = T,
      main="Histogram and p.d.f. approximation",
      xlab="weight, g")
lines(density(Mice$bw_start),lwd=2,col=4)

## (!) a box-plot of the population on the basis of sex
boxplot(bw_end~sex,data=Mice,col=c("pink","lightblue"))
title("Weight by sex (f:female, m:male)",
      ylab="weight, g",xlab="sex")

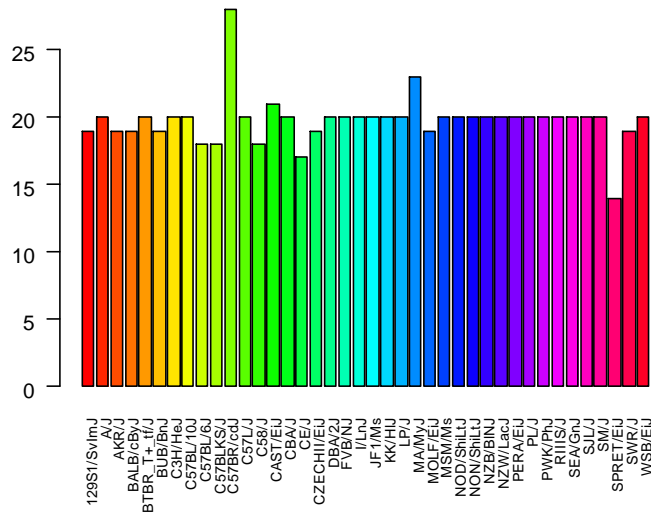
```

```

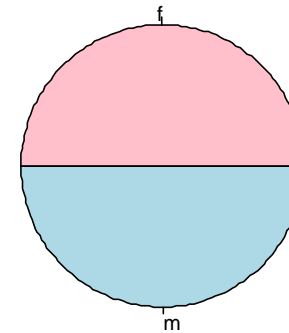
> str(Mice)
'data.frame':   790 obs. of  32 variables
 $ strain      : Factor w/ 40 levels '
 $ sex         : Factor w/ 2 levels 'f
 $ id          : int   1 2 3 368 369 37
 $ age         : int   66 66 66 72 72 7
 $ bw_start   : num   19.3 19.1 17.9 1
 $ bw_end     : num   20.5 20.8 19.8 2
 $ bw_chg     : num    1.06 1.09 1.11 1
 $ CaCl2_pref7 : num   33.3 58.3 63.3 5
 $ CaCl2_pref25 : num   66.7 47.1 55.8 5
 $ CaCl2_pref75 : num   38.8 27.9 41.9 3
 $ CaLa_pref7  : num   65.2 68 57.1 58.
 $ CaLa_pref25 : num   61.9 93.3 65.9 6
 $ CaLa_pref75 : num   45.5 23.3 73.3 4
 $ NaCl_pref25 : num   72.2 91.1 64.9 5
 $ NaCl_pref75 : num   80.2 94.3 86.6 4
 $ NaCl_pref225 : num   88.7 95.8 93.4 1
 $ NaLa_pref25 : num   79.5 97.6 91 21.
 $ NaLa_pref75 : num   86.7 97.2 91.5 3
 $ NaLa_pref225 : num   89.8 97.6 95.1 8
 $ bleeding_time : num   64 78 90 65 55 1
 $ ionized_Ca  : num    1.2 1.15 1.16 1.
 $ pH          : num    7.24 7.27 7.26 7
 $ adj_ionized_Ca : num    1.12 1.09 1.09 1
 $ total_calcium : num    2.34 2.17 2.37 2
 $ age_end     : int  116 116 108 114
 $ BMD         : num    0.0605 0.0553 0.
 $ BMC         : num    0.536 0.493 0.51
 $ lean_wt     : num    14.5 13.9 13.8 1
 $ fat_wt      : num    4.4 4.4 2.9 4.2
 $ total_wt    : num    18.9 18.3 16.7 1
 $ pct_fat     : num    23.3 24 17.4 21.
 $ pct_lean    : num    76.7 76 82.6 78.

```

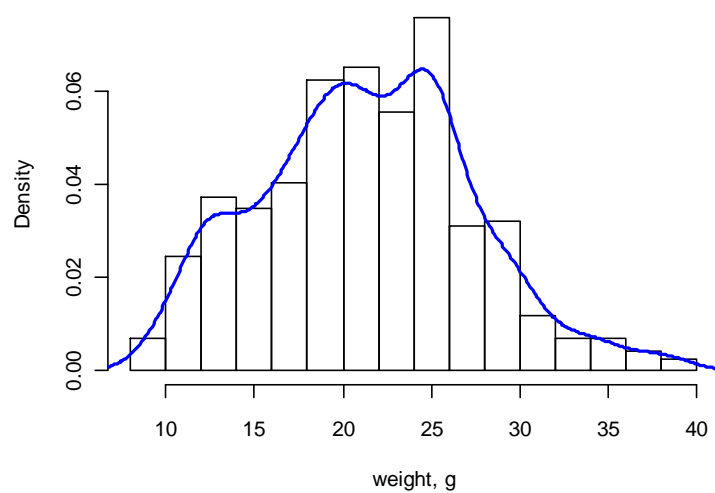
Number of mice from each strain



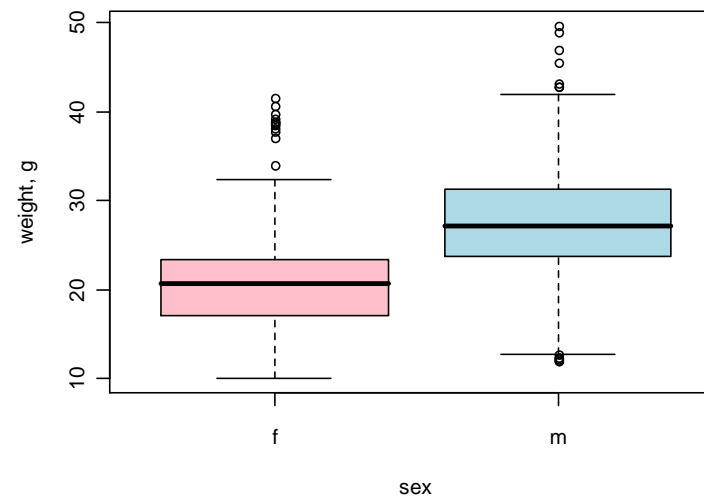
Gender composition (f:female, m:male)



Histogram and p.d.f. approximation



Weight by sex (f:female, m:male)



```

##-----
## 7.1. Descriptive statistics
##-----
str(Mice)
## summary on all the variables
summary(Mice)
## descriptive statistics on weight
bw=Mice$bw_end # to shorten typing
## let us put all information into a list
ms = list()
ms$mean = mean(bw) # average
ms$median = median(bw) # robust average estimation
ms$st.deviation = sd(bw) # measure of data variability
ms$variance = var(bw) # st.deviation ^ 2
ms$median.absolute.deviation =
  mad(bw) # robust measure of data variability
ms$min = min(bw) # min
ms$max = max(bw) # max
ms$correlation.with.initial.weight =
  cor(bw,Mice$bw_start)
ms
## see the mouse with the highest weight
Mice[which.max(bw),]
## try this
mean(Mice$bleeding_time)
## to solve
mean(Mice$bleeding_time,na.rm=T)

```

```

> ms
$mean
[1] 23.69114

$median
[1] 23.5

$st.deviation
[1] 7.068181

$variance
[1] 49.95919

$median.absolute.deviation
[1] 6.96822

$min
[1] 10

$max
[1] 49.6

$correlation.with.initial.weight
[1] 0.942258

> mean(Mice$bleeding_time)
[1] NA

> mean(Mice$bleeding_time,na.rm=T)
[1] 60.99868

```

```

##-----
## 7.2. Looking for potential effects
##-----
## Let us look for potentially linked parameters.
## To do so, we can use correlation between
## parameters as a measure of their mutual effect.

## first, transform all data to numbers use data.matrix
str(data.matrix(Mice))

## now calculate the corelation matrix
xc=cor(data.matrix(Mice), data.matrix(Mice),
        use="pairwise.complete.obs")

## create a pallete
bwr.palette =
  colorRampPalette(c("blue", "white", "red"))
## plot a heatmap
heatmap(xc, scale="none", col=bwr.palette(1000))

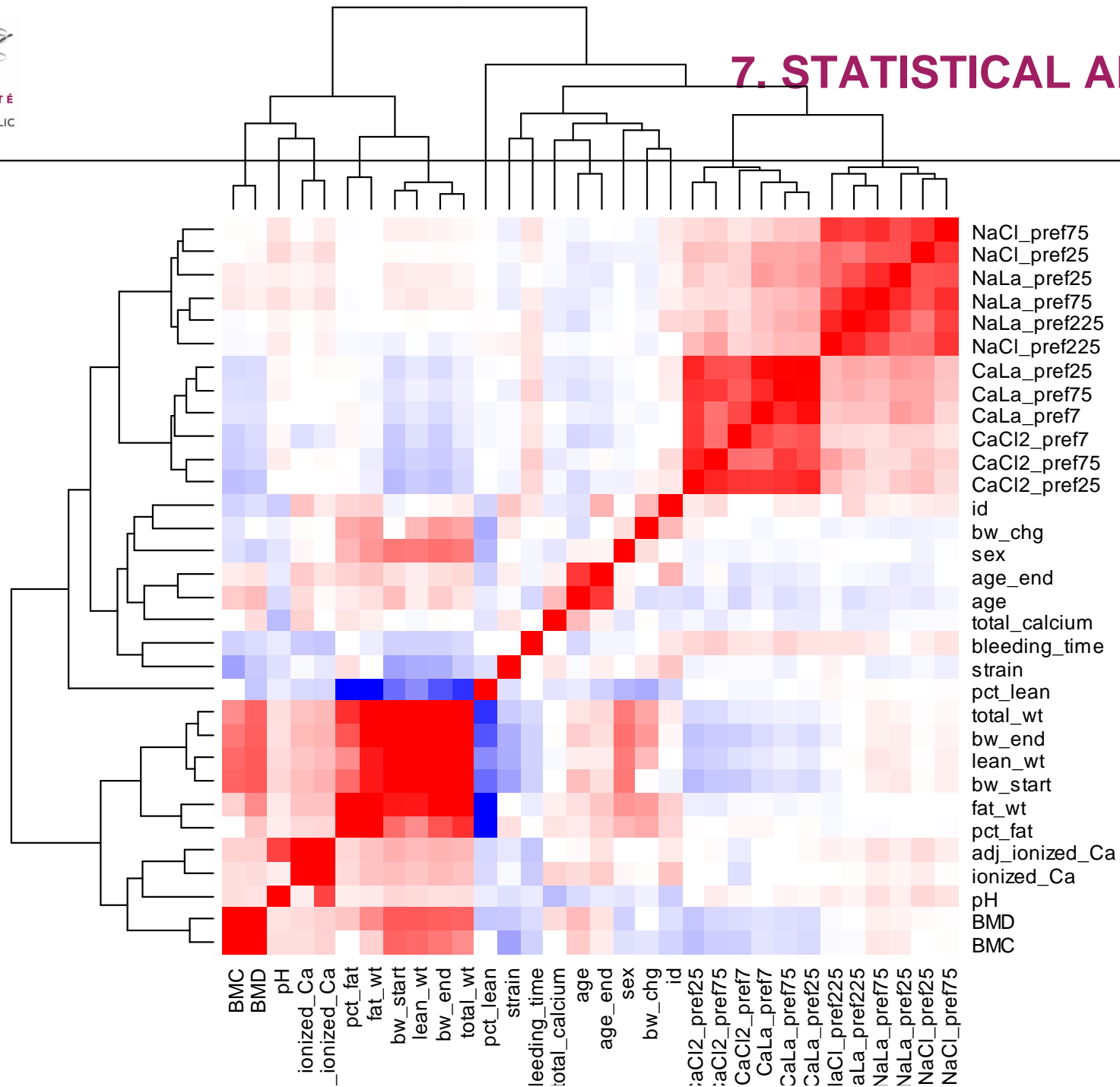
```

```

> str(data.matrix(Mice))
num [1:790, 1:32] 1 1 1 1 1
- attr(*, "dimnames")=List of 2
 ..$ : NULL
 ..$ : chr [1:32] "strain"
> xc=cor(data.matrix(Mice), data.matrix(Mice),
+        use="pairwise.complete.obs")
> bwr.palette =
+   colorRampPalette(c("blue", "white", "red"))
> heatmap(xc, scale="none", col=bwr.palette(1000))
> |

```

7. STATISTICAL ANALYSIS

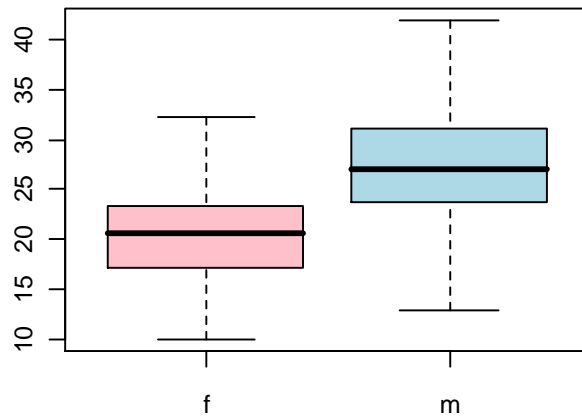


```

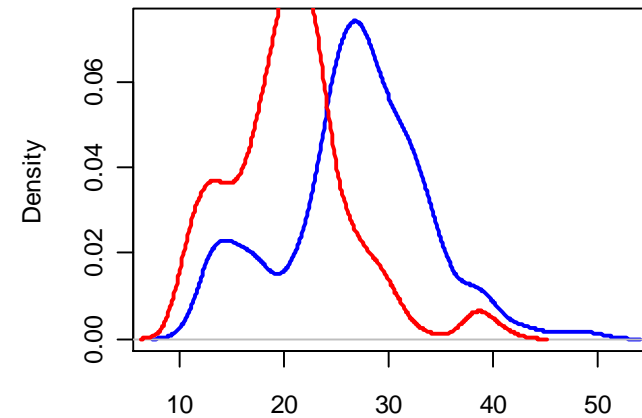
##-----
## 7.3. Statistical tests
##-----
windows ()
par (mfrow=c (2,2))
## Let us plot the distributions of the final weights
## of mice in a nice way
boxplot (bw_end~sex, data=Mice, col=c ("pink", "lightblue"), outline=F)
title ("Final body weights (g)")
plot (density (Mice$bw_end[Mice$sex=="m"]), col="blue", , lwd=2, main="")
lines (density (Mice$bw_end[Mice$sex=="f"]), col="red", lwd=2)
title ("Body weight distributions")
## And the distributions of weight increase (after experiment minus
## before experiment)
boxplot (bw_chg~sex, data=Mice, col=c ("pink", "lightblue"), outline=F)
title ("Weights increase (g)")
plot (density (Mice$bw_chg[Mice$sex=="m"]), col="blue", , lwd=2, main="")
lines (density (Mice$bw_chg[Mice$sex=="f"]), col="red", lwd=2)
title ("Distributions of weight increase")
## What can you say, looking on this pictures? :) Let's test...

```

Final body weights (g)

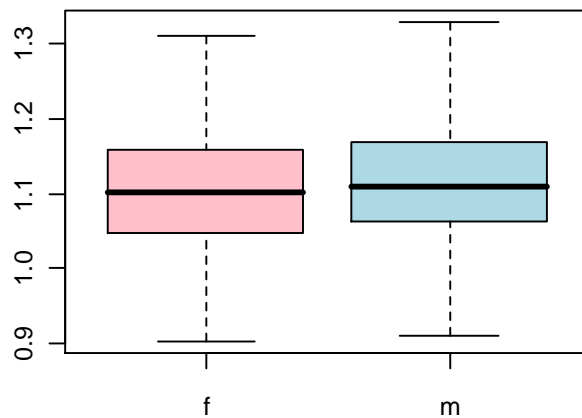


Body weight distributions

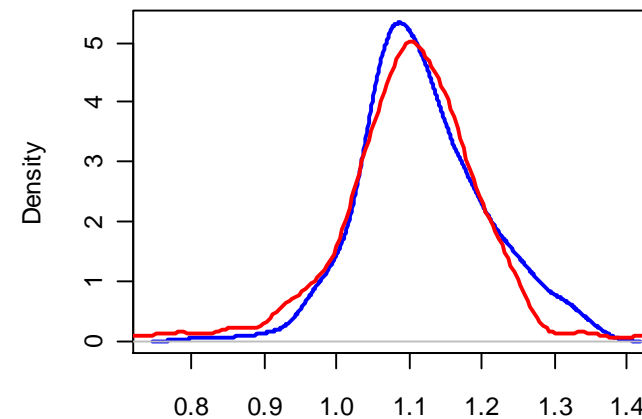


What can you say looking on these plots?

Weights increase (g)



Distributions of weight increase



N = 394 Bandwidth = 0.02154

Well, in fact almost nothing...

```
## Perform t.tests (by default - unpaired, two.sided)
t.test(Mice$bw_end[Mice$sex=="m"],Mice$bw_end[Mice$sex=="f"])
t.test(Mice$bw_chg[Mice$sex=="m"],Mice$bw_chg[Mice$sex=="f"])

## Perform Wilcoxon Rank Sum (equiv. Mann-Whitney test)
wilcox.test(Mice$bw_end[Mice$sex=="m"],Mice$bw_end[Mice$sex=="f"])
wilcox.test(Mice$bw_chg[Mice$sex=="m"],Mice$bw_chg[Mice$sex=="f"])
```

p-value = 2.2e-16

p-value = 0.0014

p-value = 2.2e-16

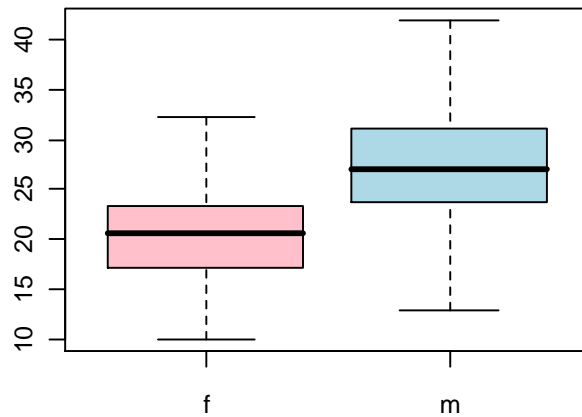
p-value = 0.029

```
> t.test(Mice$bw_chg[Mice$sex=="m"],Mice$bw_chg[Mice$sex=="f"])
```

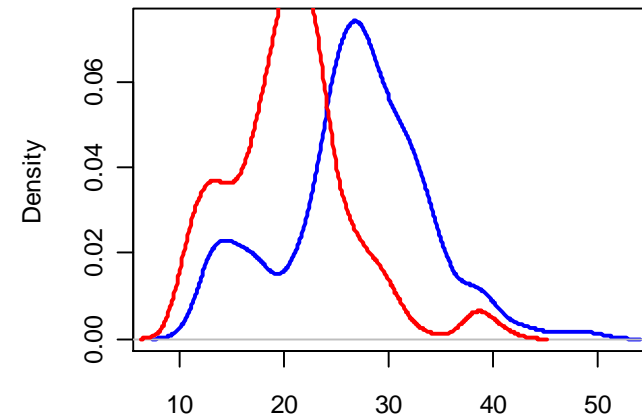
Welch Two Sample t-test

```
data: Mice$bw_chg[Mice$sex == "m"] and Mice$bw_chg[Mice$sex == $
t = 3.2067, df = 682.856, p-value = 0.001405
alternative hypothesis: true difference in means is not equal to$
95 percent confidence interval:
 0.009873477 0.041059866
sample estimates:
mean of x mean of y
 1.119401  1.093934
```

Final body weights (g)

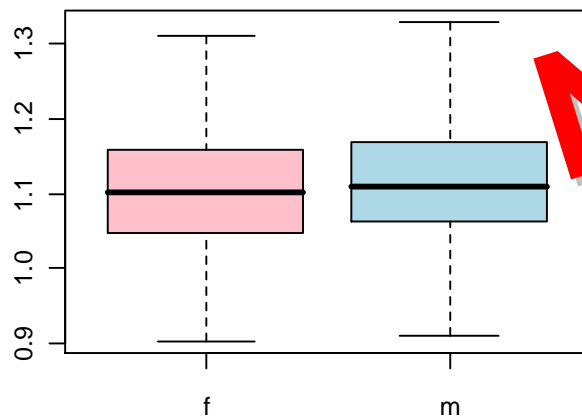


Body weight distributions

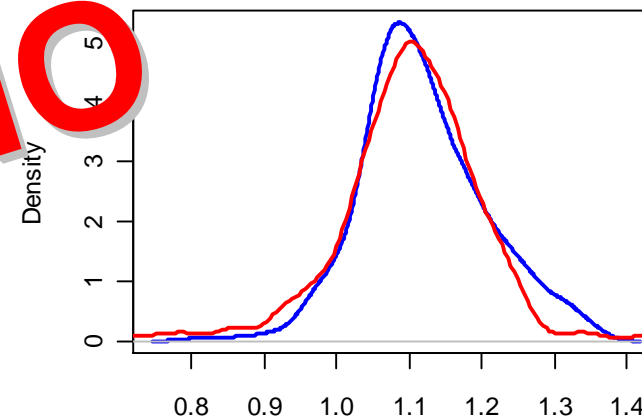


By the way, can you distinguish M and F by weight?

Weights increase (g)



Distributions of weight increase



NO

N = 394 Bandwidth = 0.02154

```

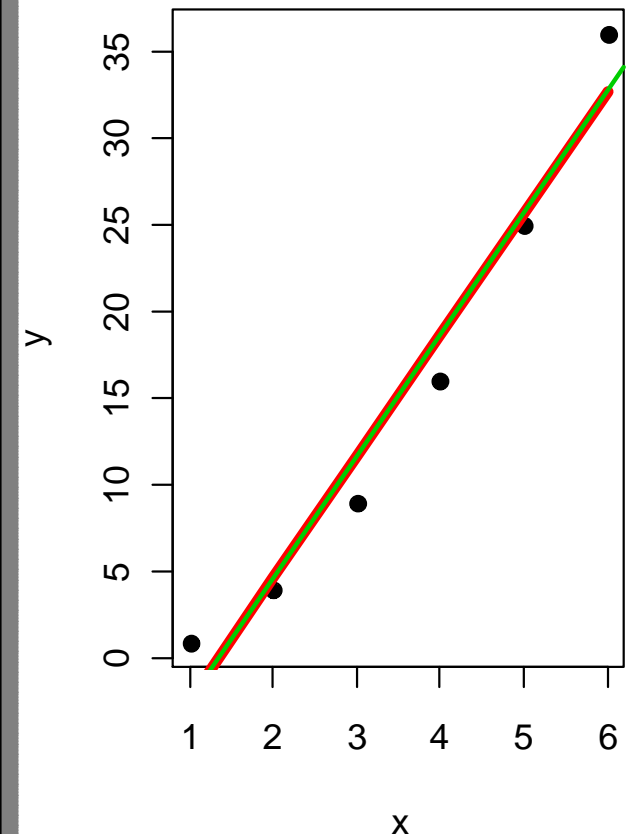
#####
# 8. LINEAR MODELS
#####
x = c(1,2,3,4,5,6) # Create ordered collection
y = x^2            # Square the elements of x
mean(y)           # Calculate arithmetic mean of y
var(y)            # Calculate sample variance

# Let us see the data
windows()
plot(y ~ x)

# Build a linear model
model=lm(y ~ x)   # Fit a linear regression model
summary(model)    # See the results
str(model)        # Look inside....

# two way of representation of the model:
lines(x,model$fitted.values,col=2,lwd=5)
abline(a=coef(model)[1],b=coef(model)[2],col=3,lwd=2)

windows()
par(mfcol=c(2,2))
plot(lm(y ~ x))
  
```



```

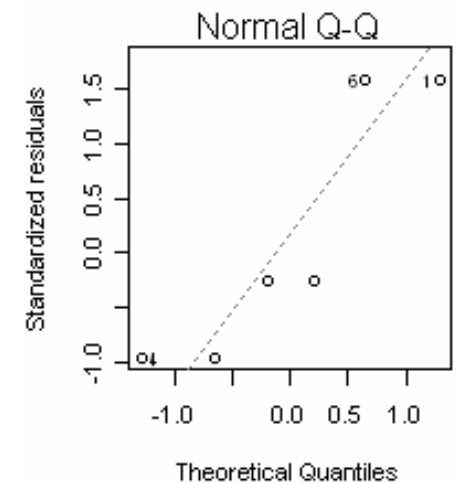
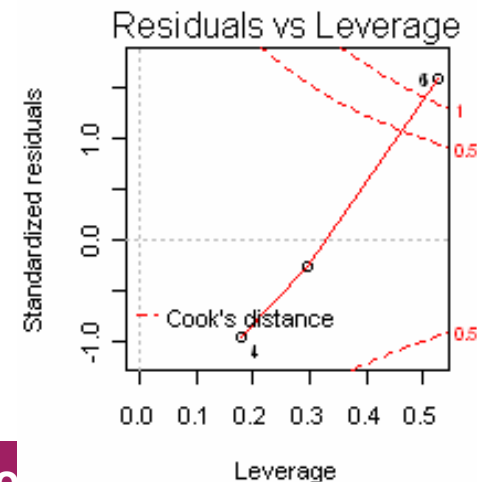
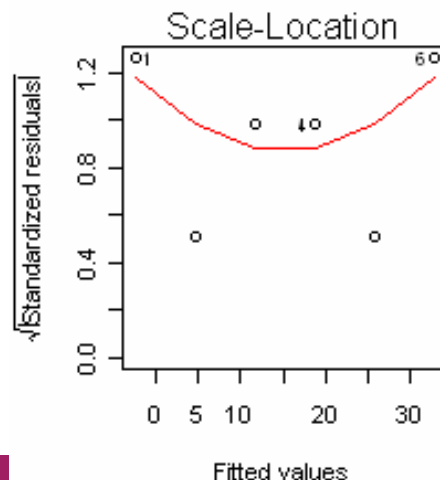
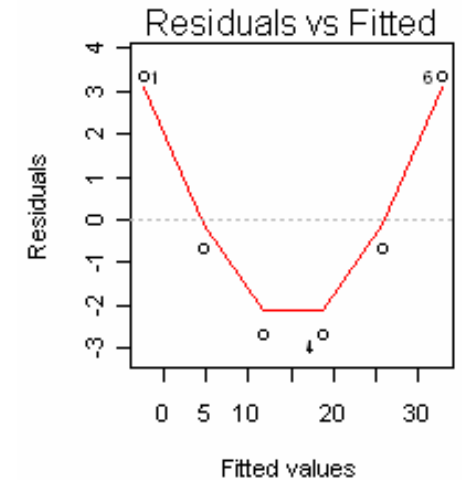
> summary(model)           # See the results

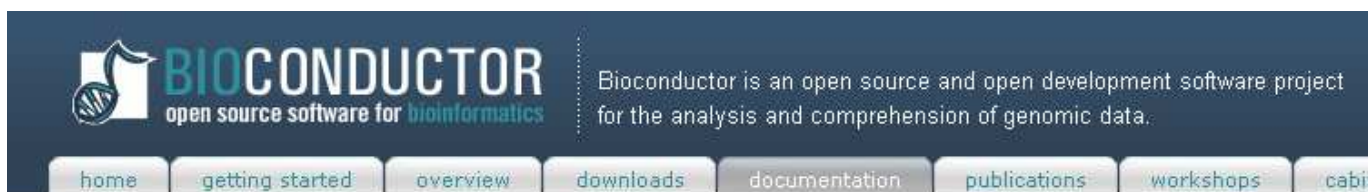
Call:
lm(formula = y ~ x)


Residuals:
    1     2     3     4     5     6 
3.3333 -0.6667 -2.6667 -2.6667 -0.6667  3.3333

Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept)  -9.3333     2.8441  -3.282  0.030453 *
x              7.0000     0.7303   9.585  0.000662 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 3.055 on 4 degrees of freedom
Multiple R-squared:  0.9583,    Adjusted R-squared:  0.9478 
F-statistic: 91.87 on 1 and 4 DF,  p-value: 0.000662
  
```






BIOCONDUCTOR

 open source software for *bioinformatics*

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Installation Instructions

Install R

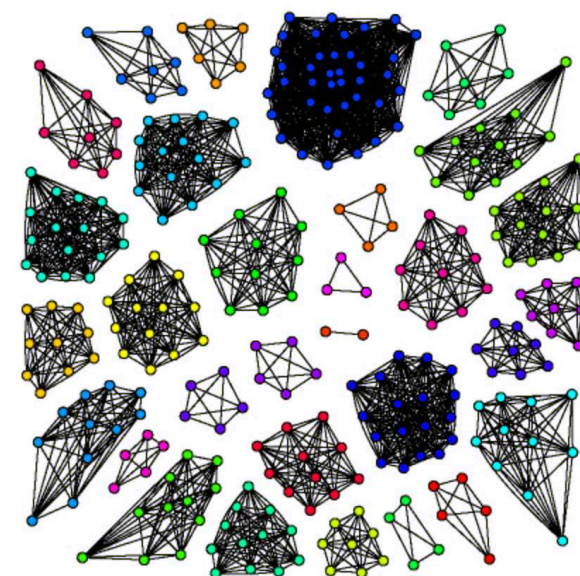
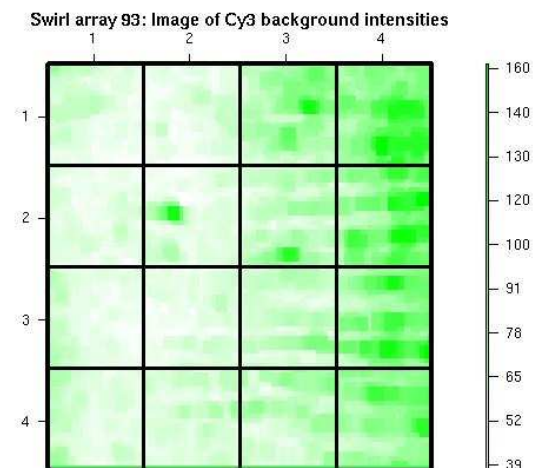
1. Download the most recent version of R from [The Comprehensive R Archive Network \(CRAN\)](#). The [R FAQ](#) and the [R Installation and Administration Manual](#) contain detailed instructions for installing R on various platforms (Linux, OS X, and Windows being the main ones).
2. Start the R program; on Windows and OS X, this will usually mean double-clicking on the R application, on UNIX-like systems, type "R" at a shell prompt.
3. As a first step with R, start the R help browser by typing "help.start()" in the R command window. For help on any function, e.g. the "mean" function, type "? mean".

Install standard Bioconductor packages

Install BioConductor packages using the `biocLite.R` installation script. In an R command window, type the following:

```
source("http://bioconductor.org/biocLite.R")
biocLite()
```

This installs the following packages: `affy`, `affydata`, `affyPLM`, `annaffy`, `annotate`, `Biobase`, `Biostrings`, `DynDoc`, `gcrma`, `genefilter`, `geneplotter`, `hgu95av2.db`, `limma`, `marray`, `matchprobes`, `multtest`, `ROC`, `vsn`, `xtable`, `affyQCReport`. After downloading and installing these packages, the script prints "Installation complete" and TRUE.



**Thank you for your
attention**

to be continued...