



LUXEMBOURG
INSTITUTE
OF **HEALTH**
RESEARCH DEDICATED TO LIFE

MaRA – an Automatic Microarray R-based Analysis Pipeline

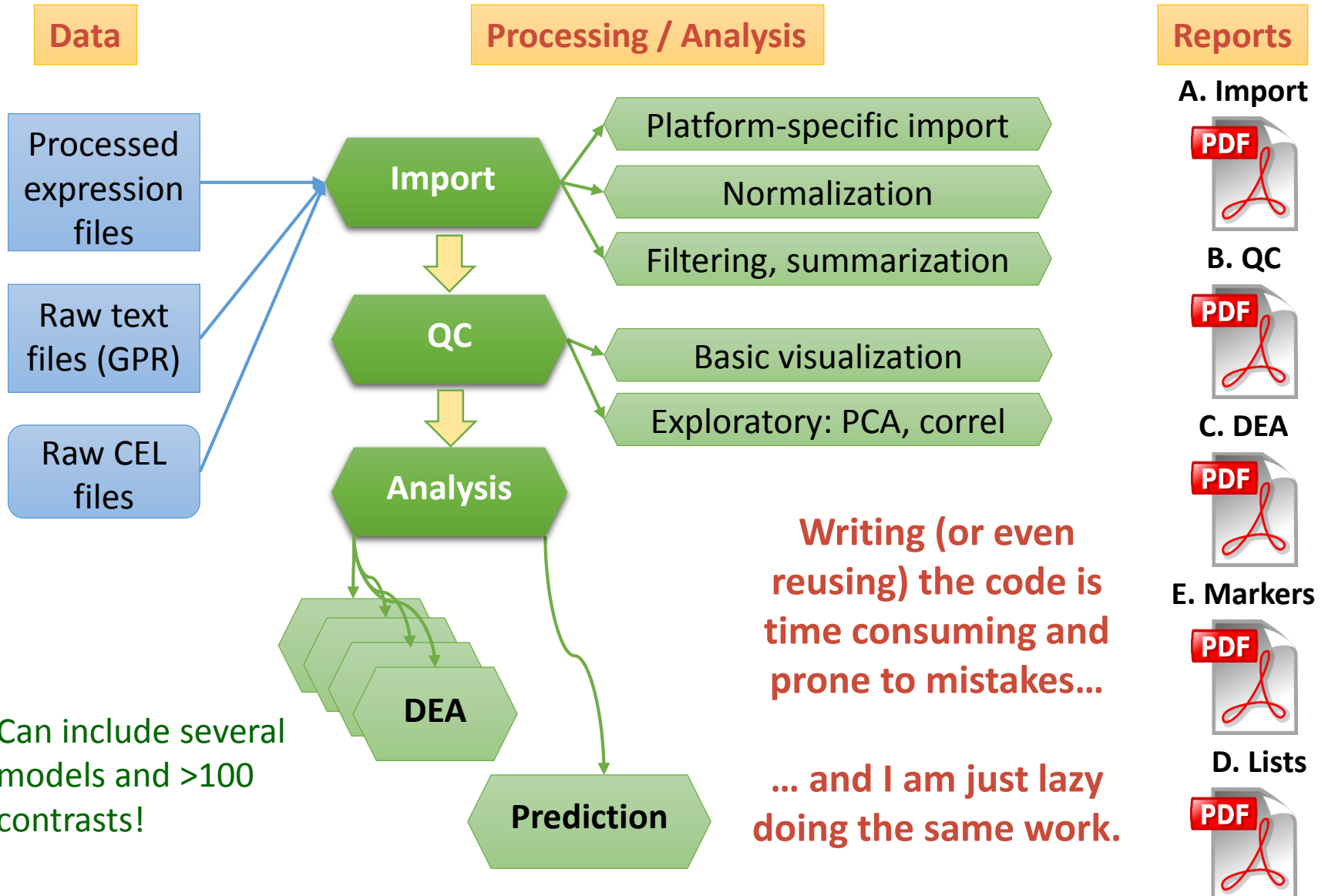
Petr Nazarov

LabMeeting, the 17th of January 2017

MaRA = a “dream” in Belarusian

- **Pipeline** in microarray analysis
- **Features** of MaRA
- **Example** – HepMirSTAT project (prof. Iris Behrman)

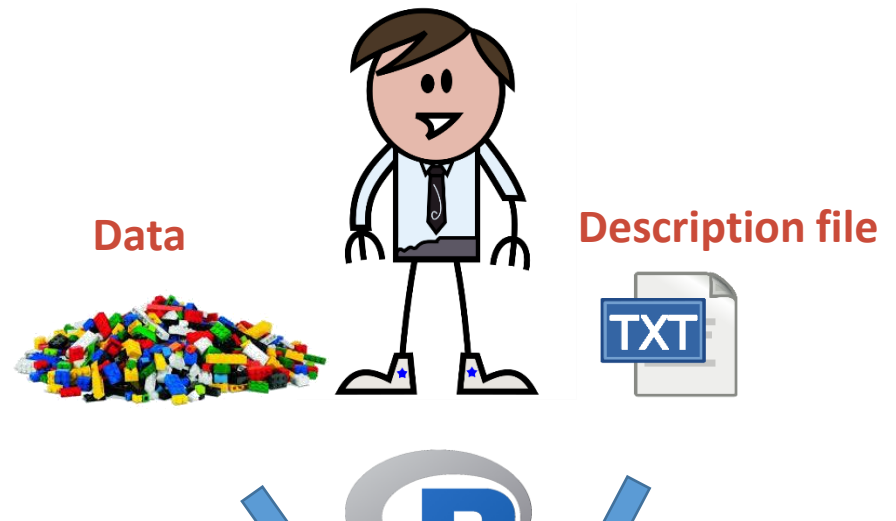
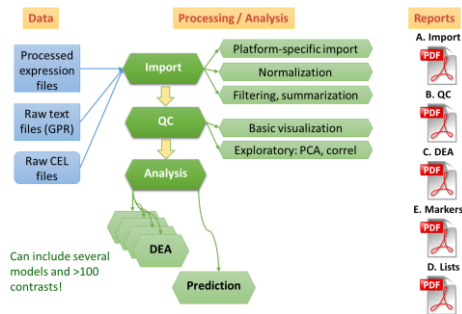
Steps of Analysis



MaRA (Microarray R-based Analysis) is an advanced flexible pipeline for automated analysis of microarray data and reporting the results.

Features

- R-based pipeline
- Scalable & flexible analysis
- Automatic processing and reporting:



Drawbacks

- No GUI (is in development)

Development

- R / Bioconductor
- Qt C++ (GUI in development)



Example of a Description File: Standard INI Configuration

[Project]

```
Title = Project Pit Ullmann
Names = P.Ullmann, E.Letellier, S.Haan
Platform = Affymetrix HuGene 2.0 ST
Description = Effect of treatments on 2 cell lines
```

[Analysis]

```
Model = Cells + Treatment + Cells*Treatment
```

```
CL18_clust.vs.ctrl = CL18, Cluster - CL18, Ctrl
```

```
CL620_clust.vs.ctrl = CL620, Cluster - CL620, Ctrl
```

```
Ctrl_18.vs.620 = CL18, Ctrl - CL620, Ctrl
```

```
Cluster_18.vs.620 = CL18, Cluster - CL620, Cluster
```

```
diffCL_Cluster.vs.Ctrl = (CL18, Cluster - CL18, Ctrl) - (CL620, Cluster - CL620, Ctrl)
```

```
ExpressionThreshold = 5
```

[Colors]

```
red = CL18, Ctrl
```

```
orange = CL18, Cluster
```

```
blue = CL620, Ctrl
```

```
cyan = CL620, Cluster
```

Example of a Description File: Standard INI Configuration

SDE genes found in contrasts can be intersected/united/excluded: &, |, !, ()

```
[Lists]
FDR = 0.01
FC = 1
list_Colon_cancer.v.healthy = Colon_HT29.vs.NCM460 & Colon_HCT116.vs.NCM460
list_Skin_cancer.v.healthy = Skin_MelJuso.vs.NHEM & Skin_A375.vs.NHEM
list_Liver_cancer.v.healthy = Liver_Hep3B.vs.PH5CH8 & Liver_Huh7.vs.PH5CH8
list_B1_cancer.v.healthy = Colon_HT29.vs.NCM460 & Colon_HCT116.vs.NCM460 & S
```

Additional analysis – looking for markers in groups of samples

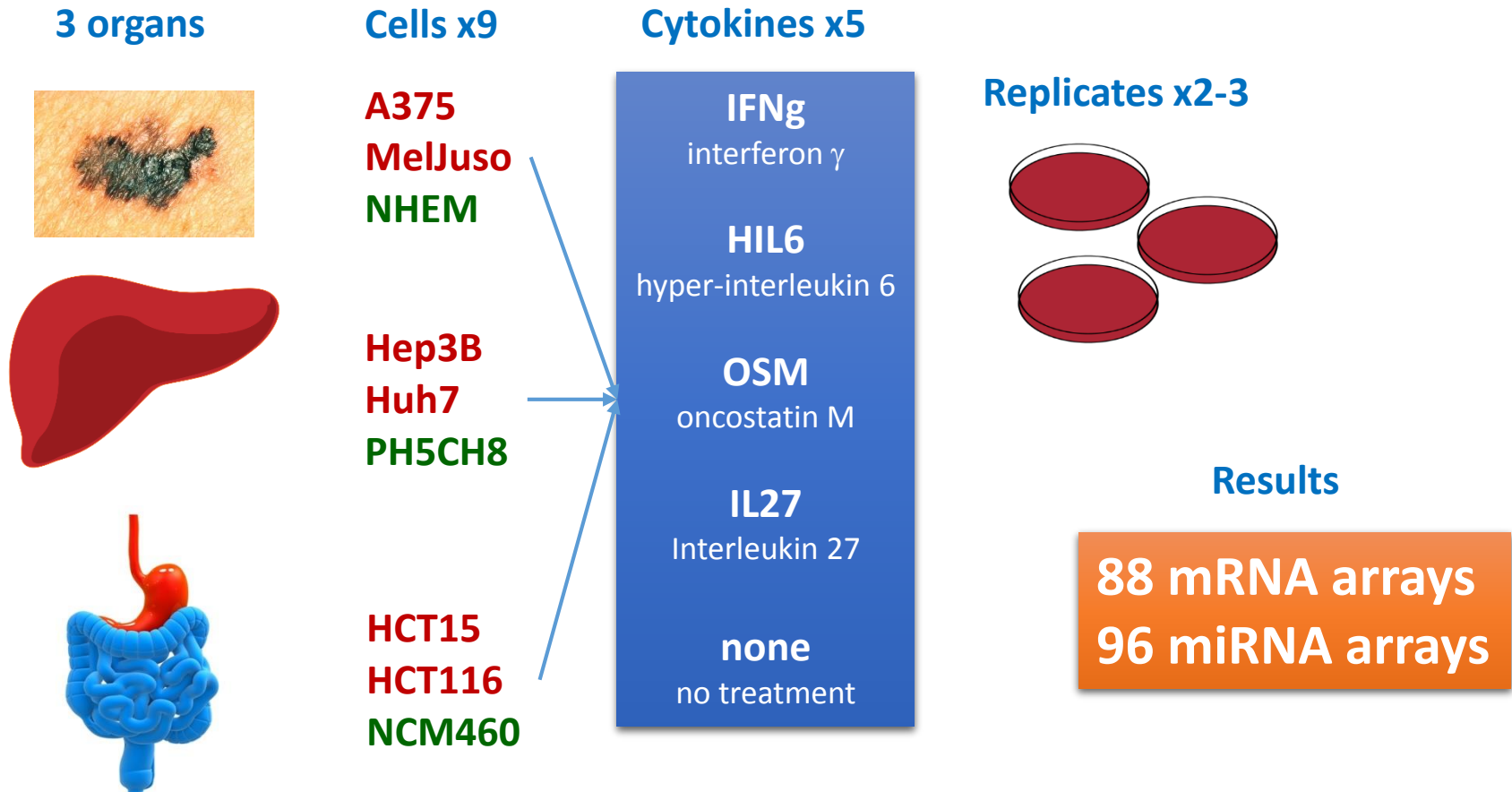
```
[Markers]
AUC = 0.99
mark_Colon = *,*,*,colon - (*,*,*,skin + *,*,*,liver)
mark_Skin = *,*,*,skin - (*,*,*,colon + *,*,*,liver)
mark_Liver = *,*,*,liver - (*,*,*,skin + *,*,*,colon)
mark_Liver.vs.Skin = *,*,*,liver - *,*,*,skin
mark_Liver.vs.Colon = *,*,*,liver - *,*,*,colon
mark_Skin.vs.Colon = *,*,*,skin - *,*,*,colon
mark_Cancer = *,*,cancer,* - *,*,normal,*
```

Example: HepMirSTAT

Project of University: cytokines in CL at mRNA & miR level

Investigation of signaling after cytokine stimulation in cell lines originated from 3 organs – skin, liver, colon.

PI - prof. Iris **BEHRMANN**, UniLu



x2 as miRNA and mRNA datasets should be investigated!

What is an effect of **each** cytokine in **each** cell line?

Can we see tissue-specific response to cytokines?

Do we have marker genes for cancer / healthy cells ?

Do we have specific genes for **each** cytokine?

Any common tendency between cytokines signaling?

Can we see cancer-specific response to cytokines?

~ 100 samples

92 comparisons

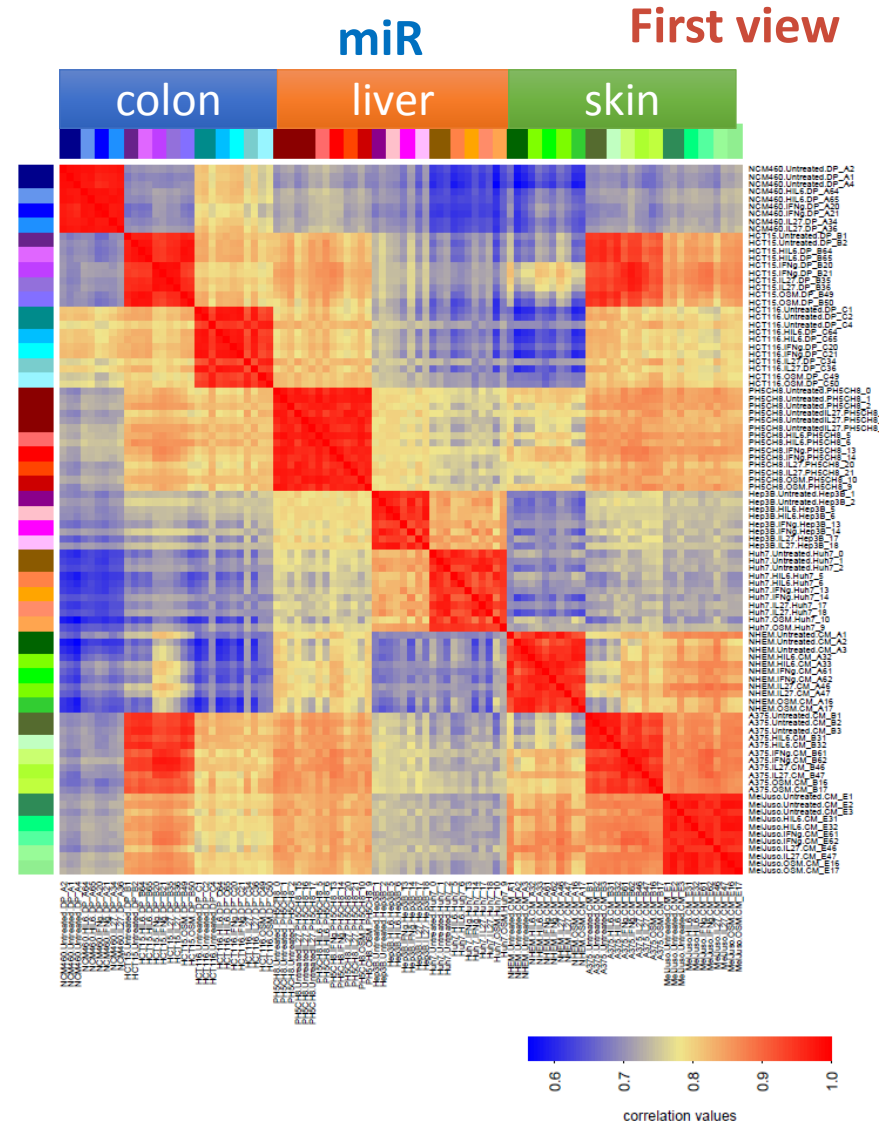
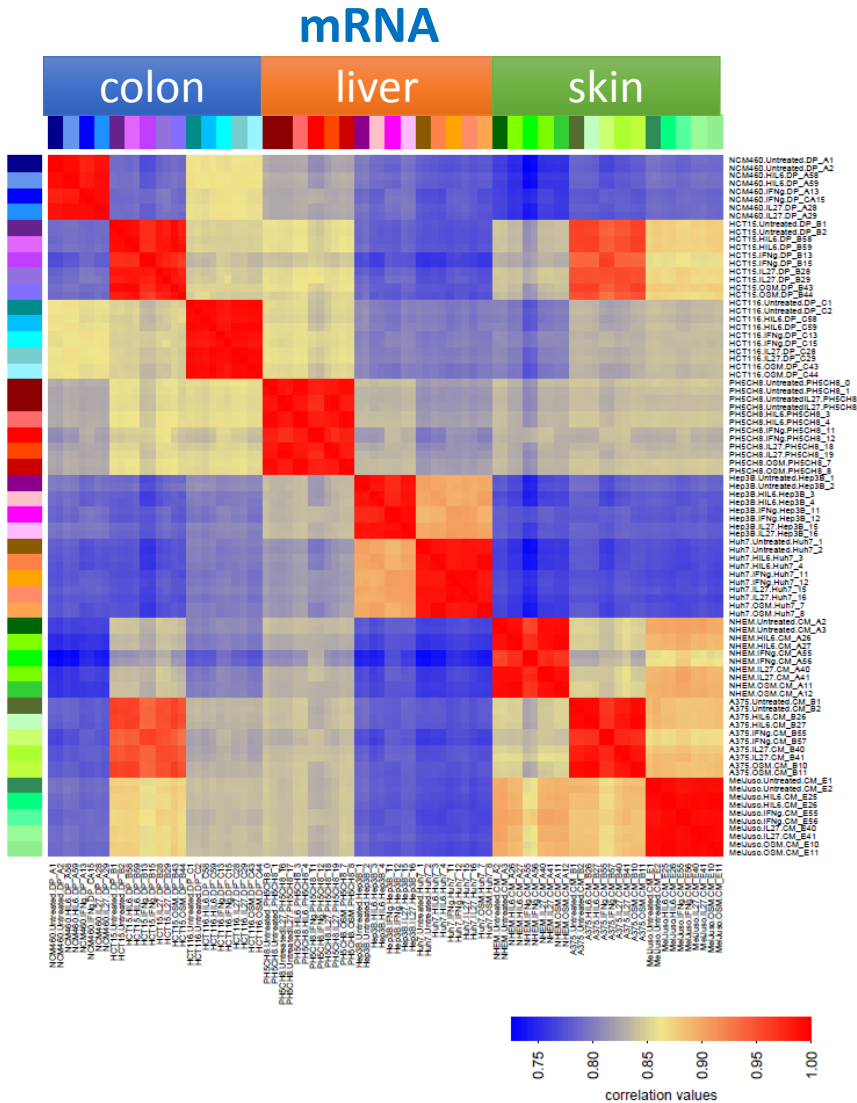
TXT

14 lists of markers

31 intersection lists

5 reports 😊

miRNA vs
mRNA vs
TargetScan



Noise in miR data is higher. Cytokine effect is much smaller than cell type effect.

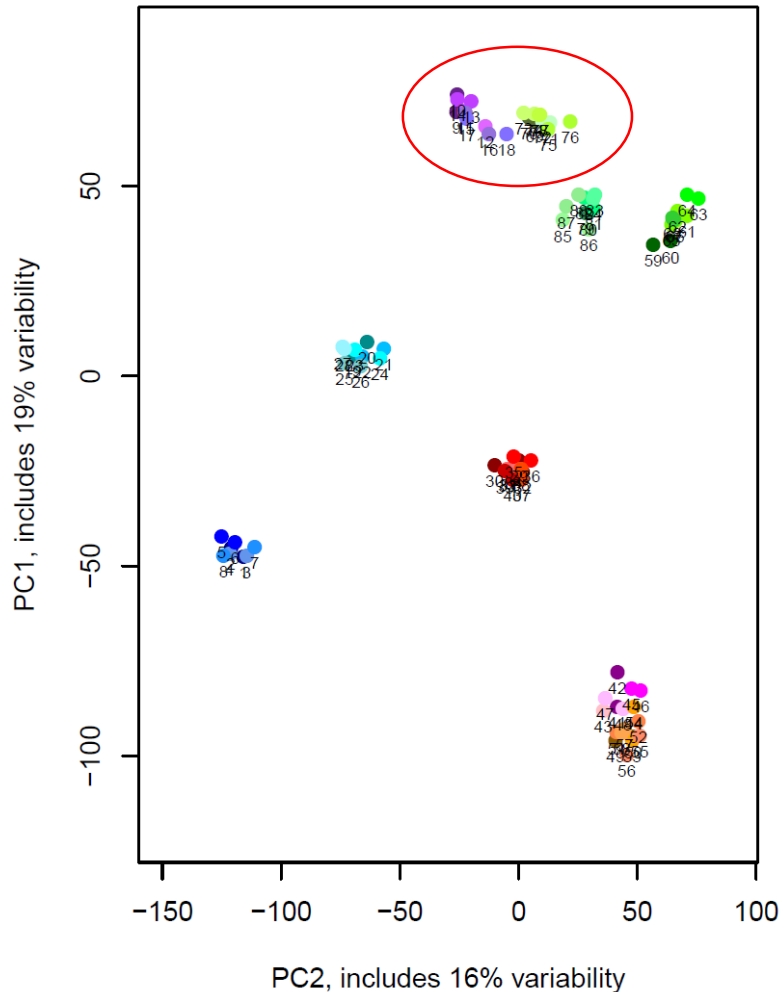
No organ grouping?

HCT15 is correlated to A375?

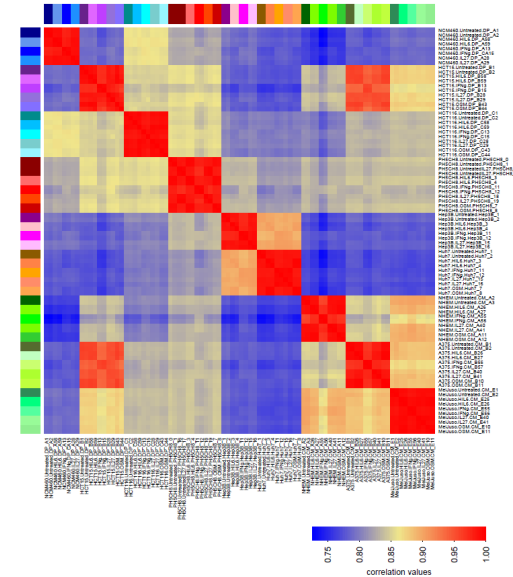
Issue?

Why is HCT15 similar to A375?

Principle component analysis (PCA) (35% variability)

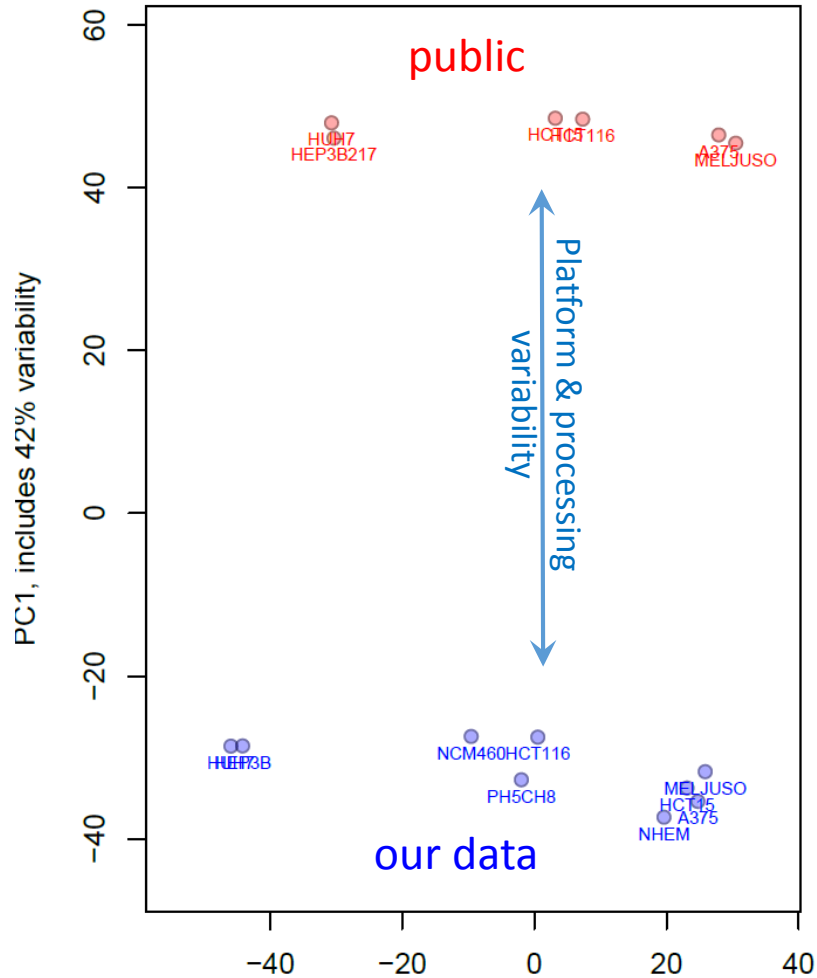


- HCT15,HIL6
- HCT15,IFNg
- HCT15,IL27
- HCT15,OSM
- HCT15,Untreated
- HCT116,HIL6
- HCT116,IFNg
- HCT116,IL27
- HCT116,OSM
- HCT116,Untreated
- NCM460,HIL6
- NCM460,IFNg
- NCM460,IL27
- NCM460,Untreated
- Hep3B,HIL6
- Hep3B,IFNg
- Hep3B,IL27
- Hep3B,Untreated
- Huh7,HIL6
- Huh7,IFNg
- Huh7,IL27
- Huh7,OSM
- Huh7,Untreated
- PH5CH8,HIL6
- PH5CH8,IFNg
- PH5CH8,IL27
- PH5CH8,OSM
- PH5CH8,Untreated
- PH5CH8,UntreatedIL27
- A375,HIL6
- A375,IFNg
- A375,IL27
- A375,OSM
- A375,Untreated
- MelJuso,HIL6
- MelJuso,IFNg
- MelJuso,IL27
- MelJuso,OSM
- MelJuso,Untreated
- NHEM,HIL6
- NHEM,IFNg
- NHEM,IL27
- NHEM,OSM

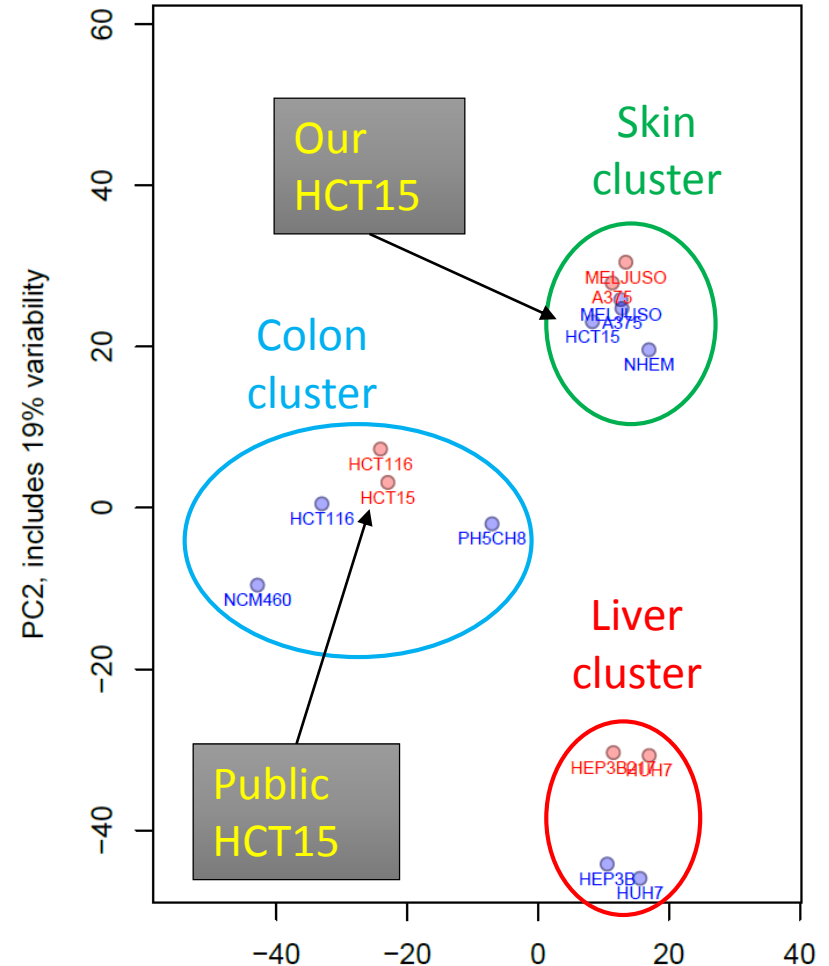


Let's have a deeper look with **Cancer Cell Line Encyclopedia (CCLE)** data

PCA 1-2
(60% variability)



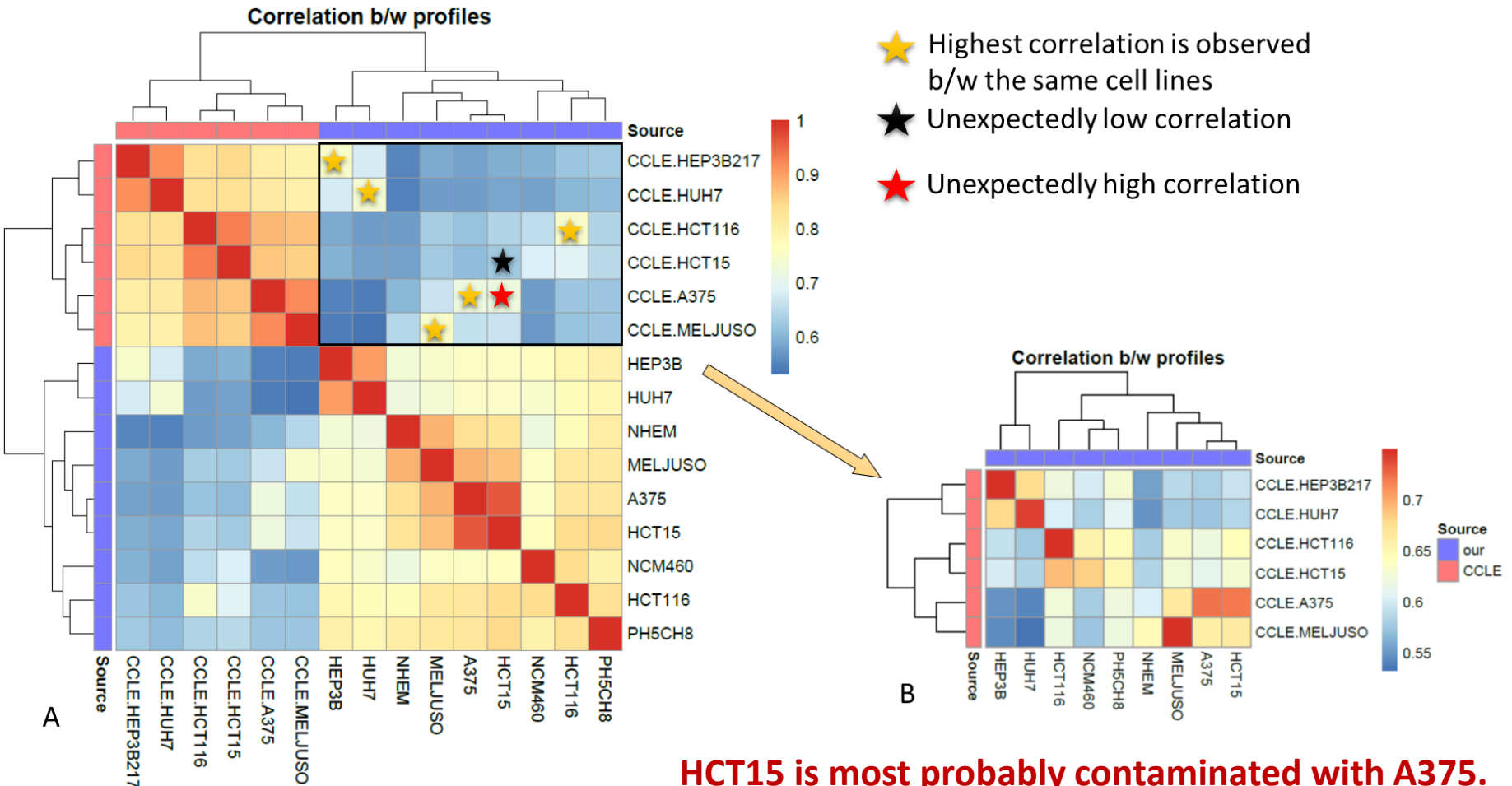
PCA 2-3
(29% variability)



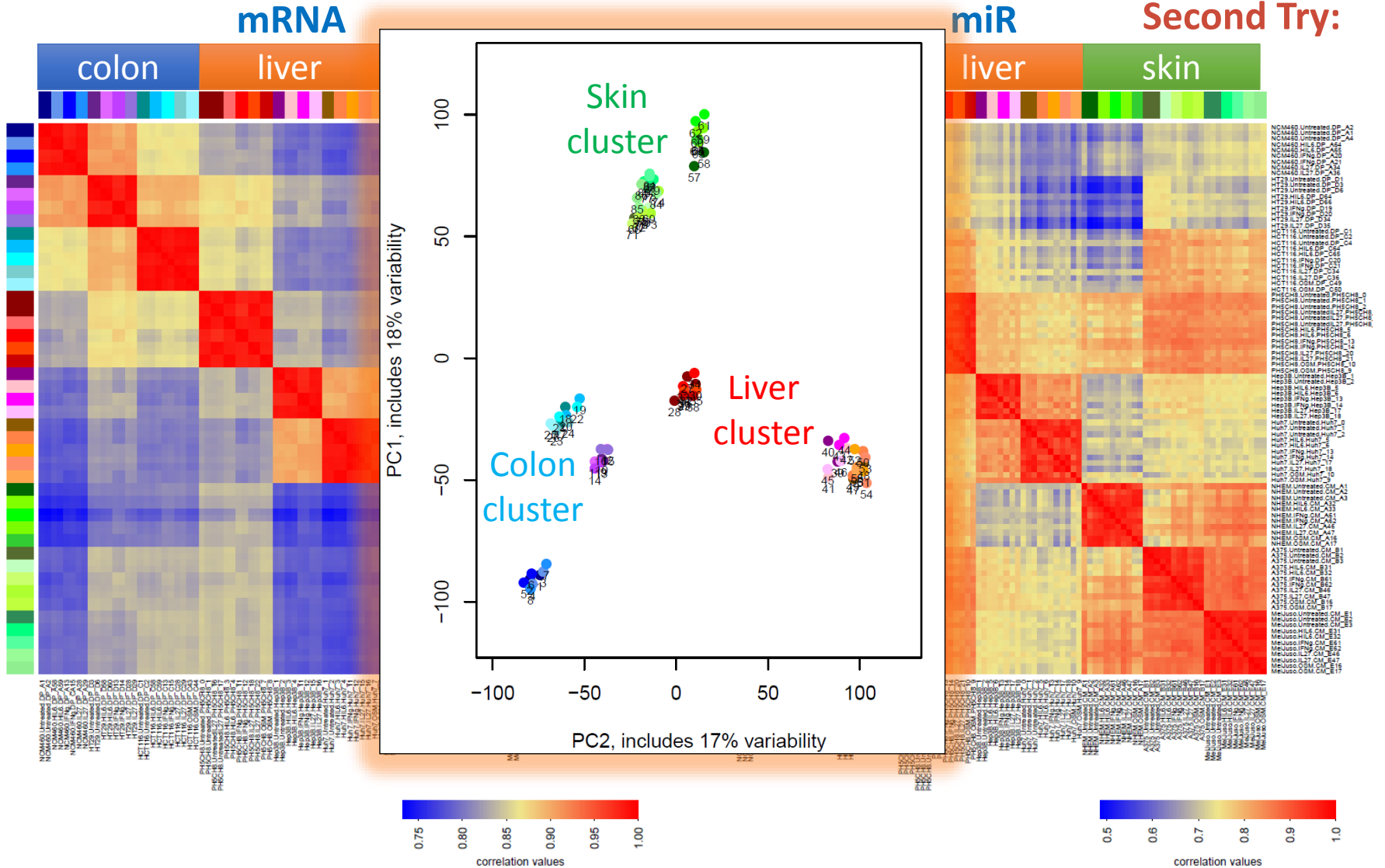
PC 1 captures b/w platform differences

PC 2-3 capture tissue differences

HCT15 is most probably contaminated by A375



**HCT15 is most probably contaminated with A375.
We replaced it with HT29**

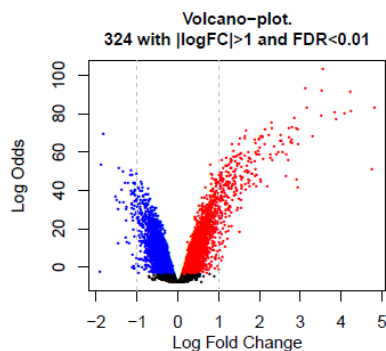
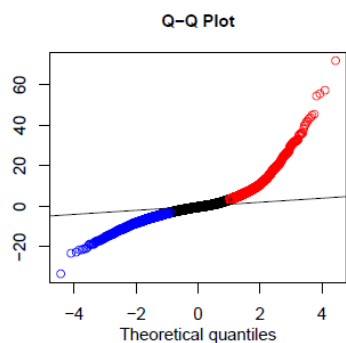


It is much better now!

A375_IFNg =
A375,IFNg - A375,Untreated

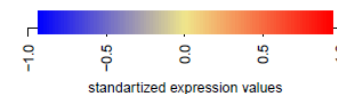
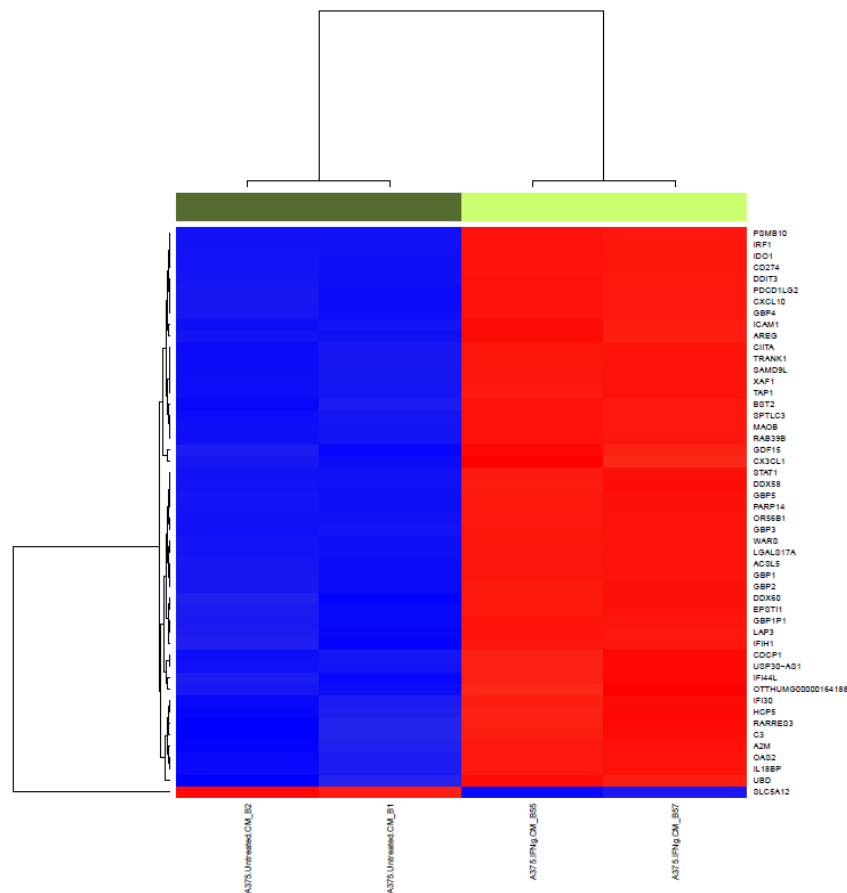
	\logFC >0	\logFC >0.5	\logFC >1	\logFC >2
FDR<0.05	8415	1826	324	52
FDR<0.01	6640	1813	324	52
FDR<0.001	5027	1768	323	52

A375_IFNg =
A375,IFNg - A375,Untreated



Top 25 probes. Look for the complete list in DE=A375_IFNg.txt

logFC	AveExpr	adj.P.Val	Gene.Symbol	gene_assignment
4.8199	5.8536	0	GBP4	NM_052941 // GBP4 // guanylate binding protein 4 // 1p22.2 // 115381 //
4.2209	5.1924	0	CD274	NM_014143 // CD274 // CD274 molecule // 9p24 // 29126 // ENST000001
3.547	5.809	0	IRF1	NM_002198 // IRF1 // interferon regulatory factor 1 // 5q31.1 // 3659 // E
4.2401	6.9842	0	GBP1	NM_002053 // GBP1 // guanylate binding protein 1, interferon-inducible
4.0819	5.2839	0	IDO1	NM_002184 // IDO1 // indoleamine 2,3-dioxygenase 1 // 8p12-p11 // 36
3.524	4.8014	0	GBP5	NM_001134486 // GBP5 // guanylate binding protein 5 // 1p22.2 // 11534
3.3889	5.7626	0	GBP1P1	NR_003133 // GBP1P1 // guanylate binding protein 1, interferon-inducib
3.8732	5.7348	0	XAF1	NM_017523 // XAF1 // XIAP associated factor 1 // 17p13.1 // 54739 // N
3.1251	9.0768	0	STAT1	NM_007315 // STAT1 // signal transducer and activator of transcription 1
3.5092	6.5973	0	GBP3	NM_018284 // GBP3 // guanylate binding protein 3 // 1p22.2 // 2635 // E
3.1604	4.8776	0	LGALS17A	NR_034156 // LGALS17A // Charcot-Leyden crystal protein pseudogen
4.7524	5.4888	0	CXCL10	NM_001585 // CXCL10 // chemokine (C-X-C motif) ligand 10 // 4q21 //
3.3013	5.6871	0	SAMD9L	NM_152703 // SAMD9L // sterile alpha motif domain containing 9-like //
2.8489	5.7948	0	CIITA	NM_000248 // CIITA // class II, major histocompatibility complex, transac
2.9338	7.1773	0	TAP1	NM_000503 // TAP1 // transporter 1, ATP-binding cassette, sub-family I
2.8952	6.464	0	ACSL5	NM_018234 // ACSL5 // acyl-CoA synthetase long-chain family membe
2.7947	4.6455	0	POCD1LG2	NM_025239 // POCD1LG2 // programmed cell death 1 ligand 2 // 9p24.2
2.7202	6.6156	0	A2M	NM_000014 // A2M // alpha-2-macroglobulin // 12p13.31 // 2 // ENST01
2.6483	6.9096	0	GBP2	NM_004120 // GBP2 // guanylate binding protein 2, interferon-inducible
2.7911	6.6802	0	BST2	NM_004335 // BST2 // bone marrow stromal cell antigen 2 // 19p13.1 // I
2.718	5.7971	0	UBD	NM_006398 // UBD // ubiquitin D // 6p21.3 // 10537 // ENST0000037701
2.7075	5.7914	0	OAS2	NM_001032731 // OAS2 // 2'-5'-oligoadenylate synthetase 2, 9971kDa
2.7813	6.4274	0	AREG	NM_001057 // AREG // amphiregulin // 4q13.3 // 374 // ENST00000264
2.5133	5.9309	0	EPST11	NM_001002284 // EPST11 // epithelial stromal interaction 1 (breast) // 13
2.2894	5.7031	0	MAOB	NM_000898 // MAOB // monoamine oxidase B // Xp11.23 // 4129 // ENS

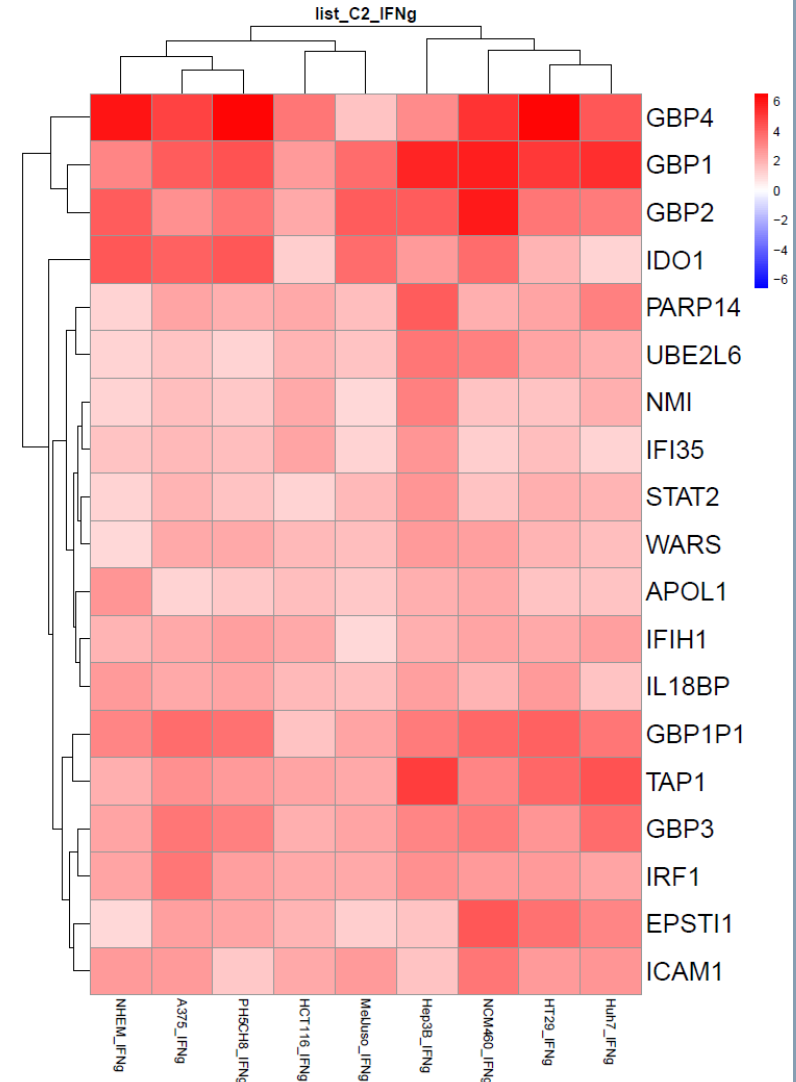


List: list_C2_IFNg

Formula: A & B & C & D & E & F & G & H & I

	List	Signif	Up	Down
A	A375_IFNg	324	263	61
B	HCT116_IFNg	100	98	2
C	HT29_IFNg	236	209	27
D	Hep3B_IFNg	221	191	30
E	Huh7_IFNg	142	138	4
F	MelJuso_IFNg	67	67	0
G	NCM460_IFNg	195	169	26
H	NHEM_IFNg	521	268	253
I	PH5CH8_IFNg	190	180	10
=>	list_C2_IFNg	19	19	0

Majority of the genes indeed comes from IFNg pathways.



mRNA

Cell line	OSM	HIL6	IL27	IFNg
MeIJuso	2	3	9	67
HCT116	0	0	25	100
PH5CH8	35	3	34	190
HT29		43	25	236
NCM460		34	79	195
Hep3B		94	102	221
A375	8	4	124	324
Huh7	134	88	106	142
NHEM	29	34	54	521

miR

Cell line	OSM	HIL6	IL27	IFNg
MeIJuso	0	1	2	13
HCT116	6	0	0	0
PH5CH8	1	0	1	6
HT29		3	0	18
NCM460		3	1	30
Hep3B		0	3	3
A375	12	1	56	44
Huh7	13	18	0	4
NHEM	10	6	3	24

FDR = $1e-2$, $|\logFC| > 1$

Common: only with IFNg stimulation – 19 mRNAs

And the rest of the interpreting is done at Uni side – we are waiting for an update meeting with them. In December they were presenting 2 posters.

Genomics Research team, LIH

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Laurent VALLAR

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Florence SERVAIS

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